



Transfer

Scalable particles:
EASY
UHPLC ↔ HPLC

Flexible

pH = 1 – 12
Temp. up to 70°C
100% aqueous
conditions

Universal

YMC-Triart
for acidic, basic and
neutral analytes

YMC-Triart

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Introduction

Chromatographers always seek to push the limits of HPLC columns to greater extremes to allow them to perform day-to-day with ever-changing pH, buffers and temperature ranges. The column for the laboratory of today must be suitable for harsh pH conditions in combination with high temperature ranges without sacrificing selectivity.

In order to meet these goals, YMC developed a novel particle technology. The revolutionary production technique provides a silica-organic hybrid stationary phase, which provides an outstandingly narrow pore size and particle size distribution. This in turn, results in low back pressures and high loadability.

Phases Overview

YMC-Triart C18

versatile applications
 first choice for
 method development
 pH-stable 1 - 12
 100% aqueous eluents 


YMC-Triart C18 ExRS

extended pH and stability
 hydrophobic substances
 positional isomers
 pH-stable 1 - 12


YMC-Triart C8

alternative to C18
 short retention time
 pH-stable 1 - 12


YMC-Triart Phenyl

aromatic compounds
 (π -electron acceptor)
 conjugated systems
 100% aqueous eluents 

YMC-Triart PFP

aromatic compounds
 (π -electron donor)
 cis-trans isomer
 polar halogenated compounds
 100% aqueous eluents 

YMC-Triart Diol-HILIC

good alternative for
 very polar compounds
 100% aqueous eluents 

YMC-Triart

Specification

	YMC-Triart C18	YMC-Triart C18 ExRS	YMC-Triart C8	YMC-Triart Phenyl	YMC-Triart PFP	YMC-Triart Diol-HILIC
Base	organic/inorganic silica					
Stationary phase	C18 (USP L1)	C18 (USP L1)	C8 (USP L7)	Phenyl (USP L11)	Pentafluorophenyl (USP L43)	Diol (USP L20)
Particle size	1.9, 3 and 5 μm					
Pore size	12 nm	8 nm	12 nm	12 nm	12 nm	12 nm
Specific surface	360 m ² /g	430 m ² /g	360 m ² /g	360 m ² /g	360 m ² /g	360 m ² /g
Carbon content	20%	25%	17%	17%	15%	—
Bonding	trifunctional					
Endcapping	multi-stage	multi-stage	multi-stage	multi-stage	none	none
pH range	1 ~ 12	1 ~ 12	1 ~ 12	1 ~ 10	1 ~ 8	2 ~ 10
Temp. range	pH 1-7: 70 °C, pH 7-12: 50 °C	pH 1-7: 70 °C, pH 7-12: 50 °C	pH 1-7: 70 °C, pH 7-12: 50 °C	50 °C	50 °C	50 °C
100% aqueous eluents	✓	✗	✗	✓	✓	✓

Particle technology

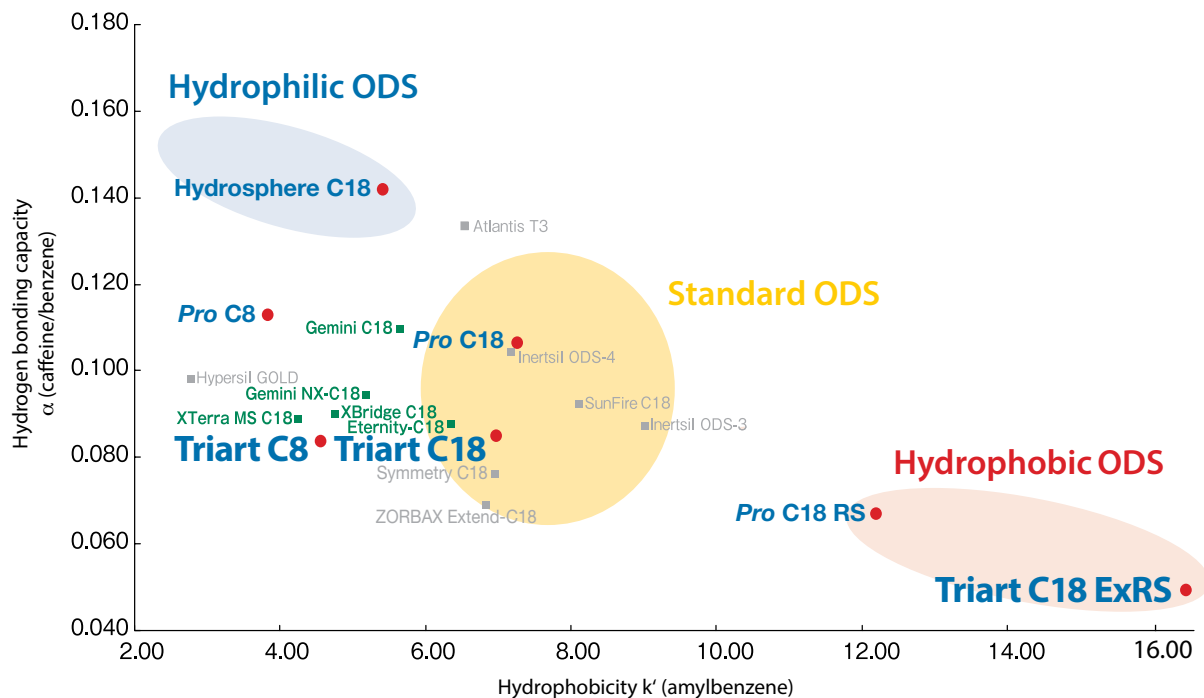
YMC-Triart is a versatile material prepared using tightly controlled particle formation technology which has been adapted from micro-reactor technology. This recently developed production process results in exceptionally narrow particle and pore size distributions.

With YMC-Triart, challenging pH and high temperature conditions are no longer a limitation to the day-to-day work in laboratories. Most importantly, due to its unique particle composition, a balanced hydrophobicity and silanol activity are achieved which makes YMC-Triart a “First Choice” column in method development.



YMC-Triart

“First choice” column for method development



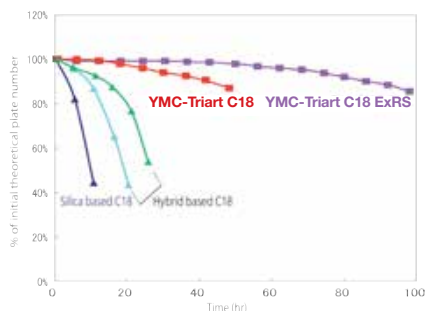
Conventional hybrid silica-based ODS columns tend to be less hydrophobic than silica-based columns. YMC-Triart C18 has a higher carbon load, giving it a hydrophobicity comparable to that of standard ODS columns, thereby making it a “versatile first-choice” column for method development.

On the other hand, YMC-Triart C18 ExRS has been designed to provide contrastingly different separation characteristics!

pH & Temperature

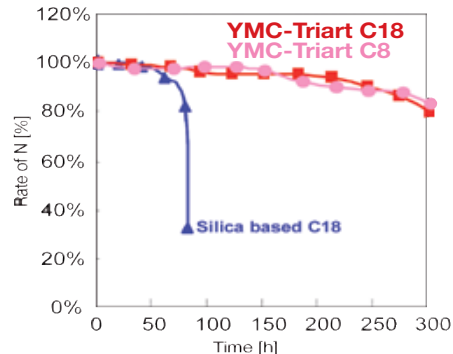
Versatile wide pH stability

Phosphate buffer (pH 11.5, 40 °C)*



Column: 5 μ m, 150 x 4.6 mm ID
 Part No.: TA12S05-1546WT
 Eluent: 50 mM K_2HPO_4 - K_3PO_4 (pH 11.5) / methanol (90/10)
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Sample: benzyl alcohol

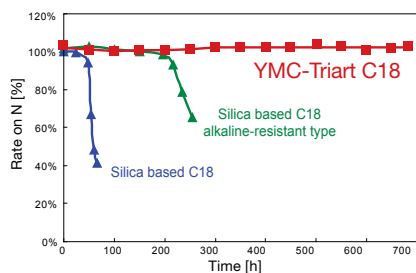
Triethylamine (pH 11.5, 40 °C)*



Column: 5 μ m, 150 x 4.6 mm ID
 Part No.: TA12S05-1546WT
 Eluent: 50 mM triethylamine (pH 11.5) / methanol (90/10)
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Sample: benzyl alcohol

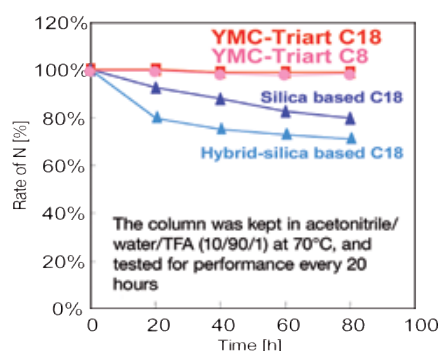
Stability at high temperature

pH 6.9, 70 °C*



Column: 5 μ m, 50 x 2.0 mm ID
 Part No.: TA12S05-0502WT
 Eluent: 20mM KH_2PO_4 - K_2HPO_4 (pH6.9)/acetonitrile(90/10)
 Flow rate: 0.2 mL/min
 Temperature: 70 °C
 Sample: phenol

pH 1, 70 °C*



Column: 5 μ m, 50 x 2.0 mm ID
 Part No.: TA12S05-0502WT
 Eluent: acetonitrile / water (60/40)
 Flow rate: 0.2 mL/min
 Temperature: 37 °C
 Sample: butyl benzoate

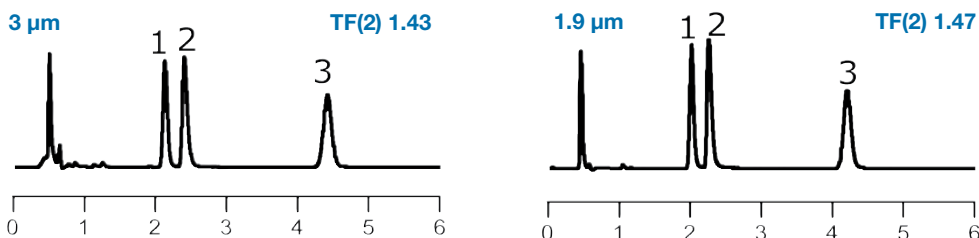
YMC-Triart phases show great chemical stability due to the highly developed hybrid-silica matrix. Even under high pH or high temperature conditions, the lifetime of YMC-Triart phases is more than 10x greater than conventional reversed phase columns.

Transfer HPLC ↔ UHPLC

Secure your method transfer!

Differences in selectivity, retention time, and also peak shapes between different particle sizes of commercially available C18 phases in the same brand (or an alternative as recommended by its manufacturer) have been observed.

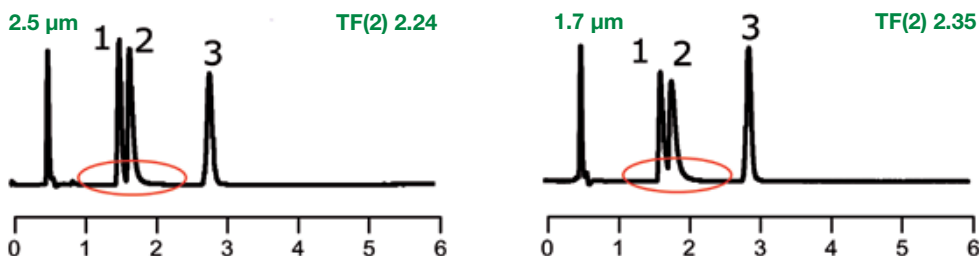
YMC-Triart C18



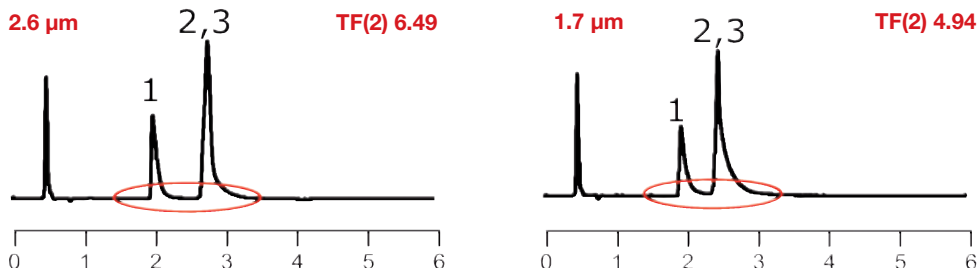
YMC has addressed this issue of method transfer. YMC-Triart columns show identical selectivity and excellent peak shapes for basic compounds for all 3.0 µm to 1.9 µm particle sizes. It allows predictable scale up from UHPLC to conventional HPLC and even to semi-preparative LC, and vice versa.

Case Studies**

X-Bridge BEH C18 and Acquity UPLC BEH C18



Kinetex™ C18



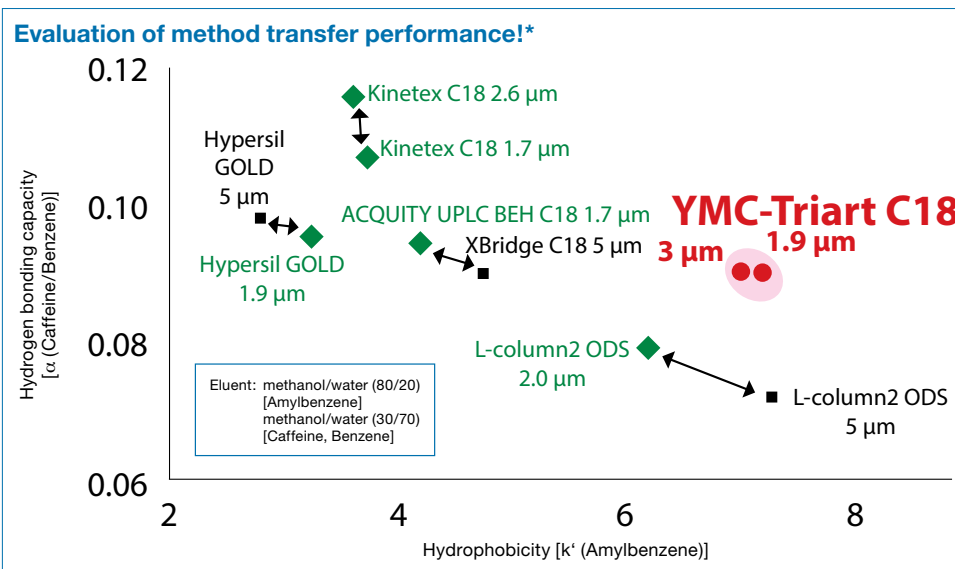
Kinetex™ C18 columns show significant peak tailing and have limited scalability due to lack of larger particle sizes.

Column: 50 x 2.0 mm ID or 2.1 mm ID
 Eluent: 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.9) / acetonitrile (65/35)
 Temperature: 40 °C
 Flow rate: 0.2 mL/min
 Detection: UV at 235 nm

1. Chlorpheniramine (basic)
 2. Dextromethorphan (basic)
 3. Propyl paraben (internal standard)

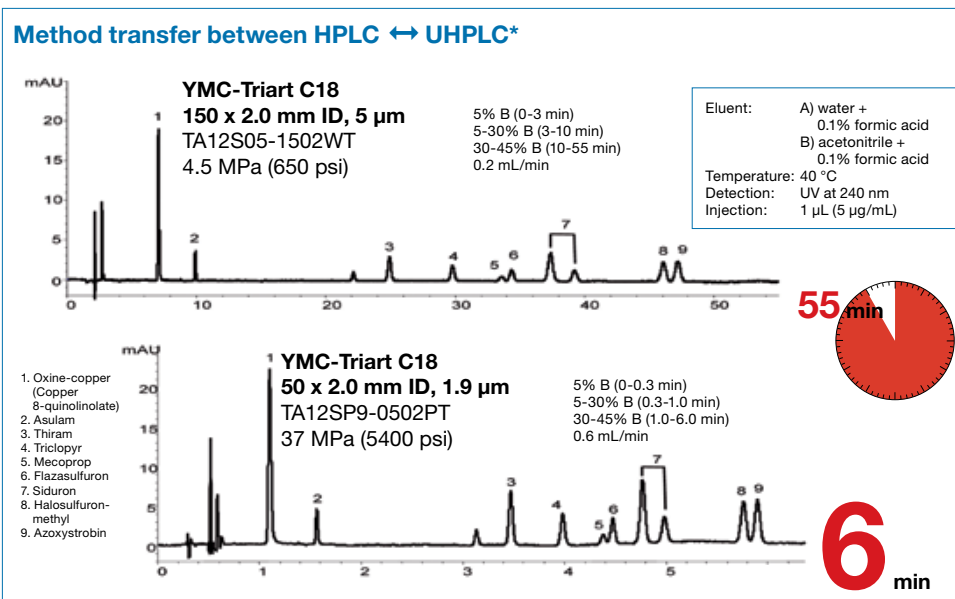
** These observations might not be representative for all applications.

Transfer HPLC ↔ UHPLC



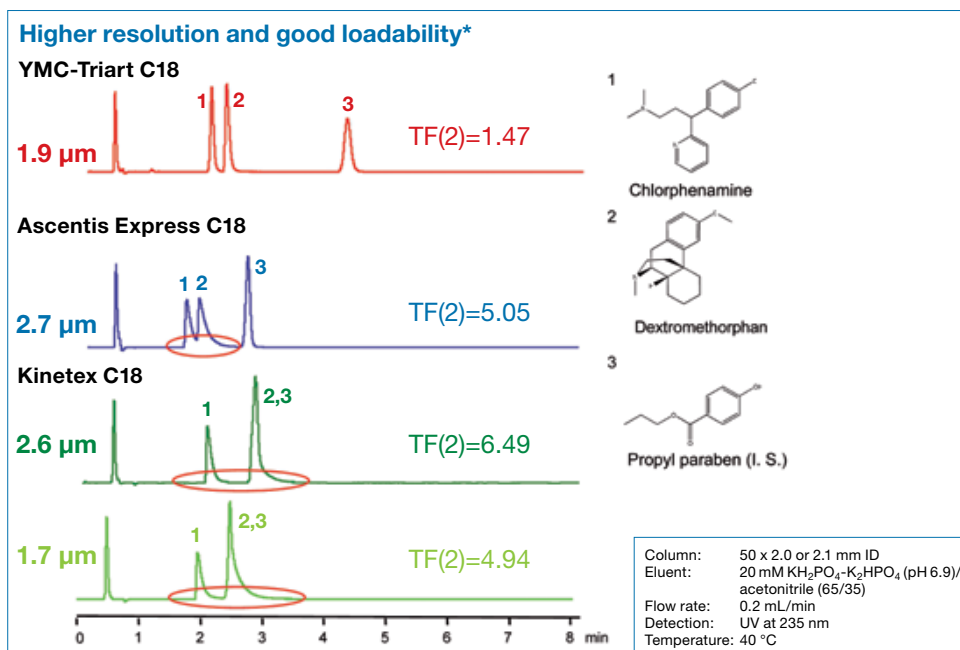
With the introduction of UHPLC, sub-2 μm particles became necessary. Therefore, smaller particles have been added to existing column lines. Consequently, sub-2 μm particles may exhibit differences in chromatographic performance.

By introducing YMC-Triart, YMC provides matching chromatographic behaviour for all particles sizes!

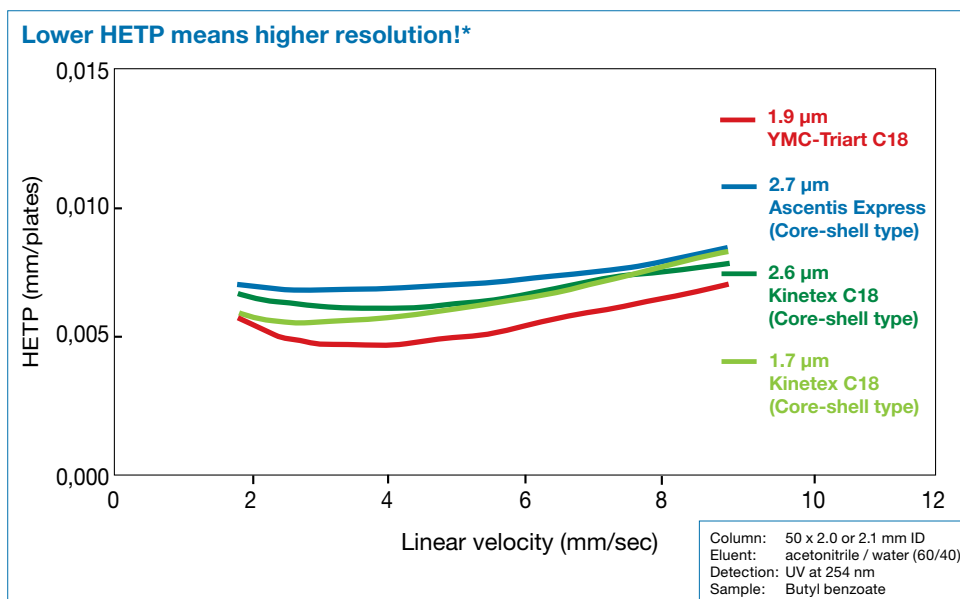


When transferring the 55 min HPLC method to UHPLC scale, the resolution remains the same although the separation time is reduced to only 6 min.

UHPLC



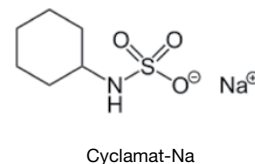
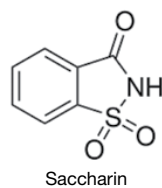
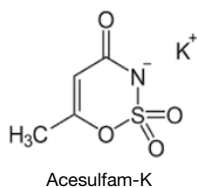
YMC-Triart C18 shows superior resolution to the other three phases thereby allowing considerable higher loadability.



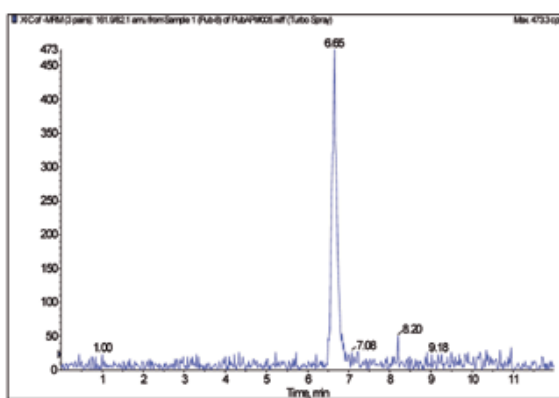
YMC-Triart C18 always shows the lowest HETP compared to the three Core-Shell products over the range of linear velocity applied.

UHPLC & MS

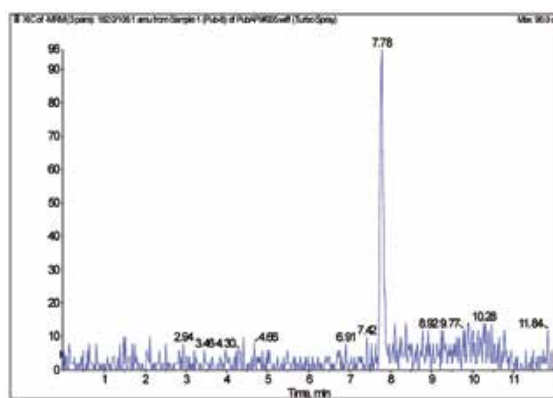
Determination of Artificial Sweeteners with LC-MS/MS



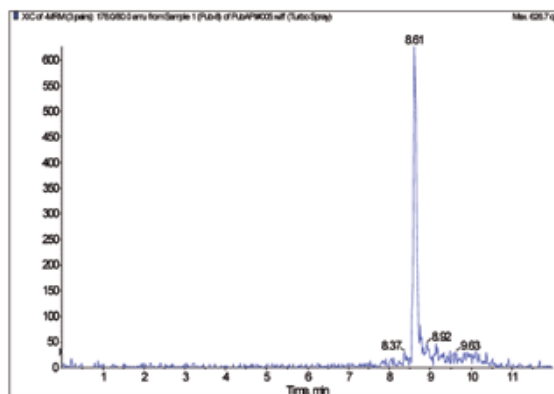
→ Non-biological markers of wastewater entries in ground and surface water



Extracted Ion Chromatogram (XIC) of Acesulfam-K, 0.1 µg/L



Extracted Ion Chromatogram (XIC) of Saccharin, 0.1 µg/L



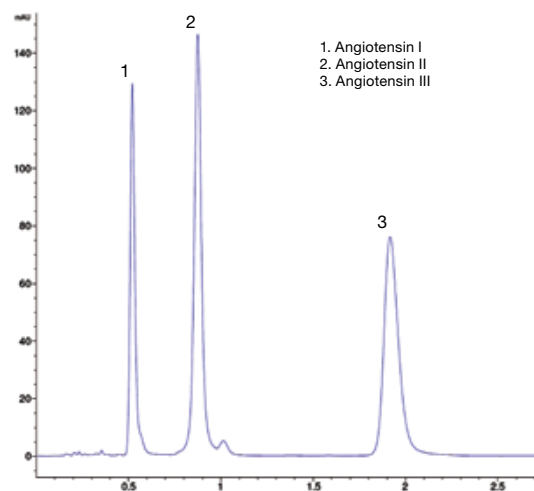
Extracted Ion Chromatogram (XIC) of Cyclamat-Na, 0.1 µg/L

Column: YMC-Triart C18 (12 nm, 1.9 µm) 100 x 3.0 mm ID
 Part No.: TA12SP9-1003PT
 LC-System: Agilent 1100 HPLC system and CTC Analytics HTC-Pal Autosampler
 MS/MS System: Applied Biosystems MDS Sciex API 4000, ESI negative
 Temperature: 35°C
 Flow rate: 0.3 mL/min
 Injection: 40 µL, direct injection
 Eluent: A: H₂O (containing 10 mmol NH₄ formate)
 B: MeOH (containing 10 mmol NH₄ formate)
 Gradient: Time 0 6.0 6.1 12.0
 % B 2 75 2 2

by courtesy of: Thomas Class, Sandro Jooß
 PTRL Europe, Helmholtzstraße 22, Science Park I, D-89081 Ulm

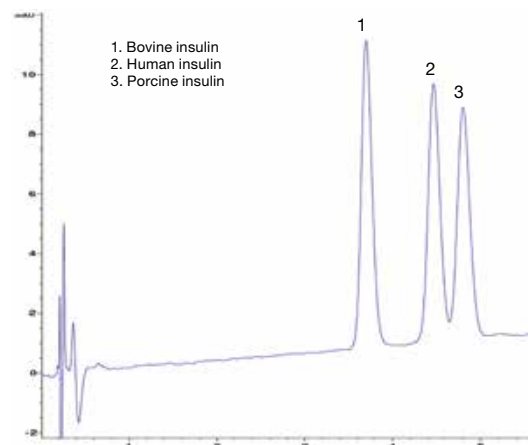
UHPLC

Angiotensin I, II and III



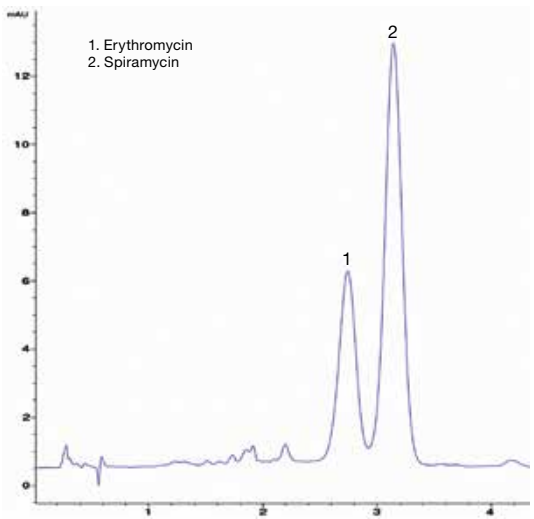
Column: YMC-Triart C18 (12 nm, 1.9 μ m) 50 x 2.0 mm ID
Part No.: TA12SP9-0502PT
Eluent: 20 mM KH_2PO_4 + K_2HPO_4 (pH 7.9) / acetonitrile (22/78)
Flow rate: 0.7 mL/min
Detection: UV at 220 nm
Pressure: 720 bar
Injection: 0.5 μ L
Temperature: 40 $^\circ\text{C}$

Insulin



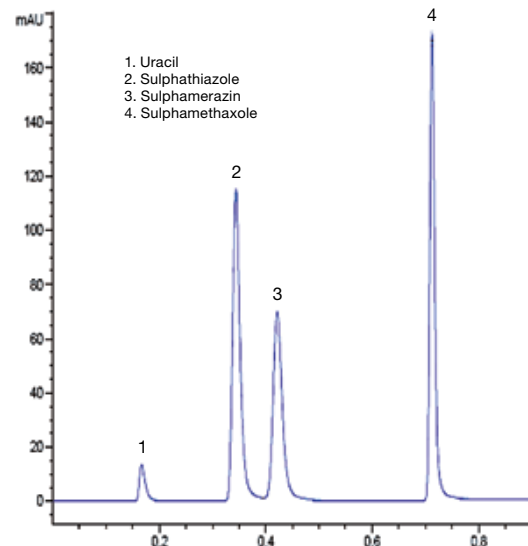
Column: YMC-Triart C18 (12 nm, 1.9 μ m) 50 x 2.0 mm ID
Part No.: TA12SP9-0502PT
Eluent: A) H_2O + 0.1% TFA
B) acetonitrile + 0.1% TFA
Gradient: 30% B (0 min); 30-32% B (0-5 min); 32% B (55 min)
Flow rate: 0.6 mL/min
Detection: UV at 220 nm
Pressure: 611 bar
Injection: 0.5 μ L
Temperature: 30 $^\circ\text{C}$

Macrolide antibiotics



Column: YMC-Triart C18 (12 nm, 1.9 μ m) 50 x 2.0 mm ID
Part No.: TA12SP9-0502PT
Eluent: A) 20 mM K_2HPO_4 + 20 mM KH_2PO_4 (pH 7.9)
B) acetonitrile
Gradient: 60% B (0.5 min); 60-70% B (0.5-1.5 min); 70% B (3.5 min)
Flow rate: 0.45 mL/min
Detection: UV at 210 nm
Pressure: 520 bar
Injection: 1 μ L
Temperature: 50 $^\circ\text{C}$

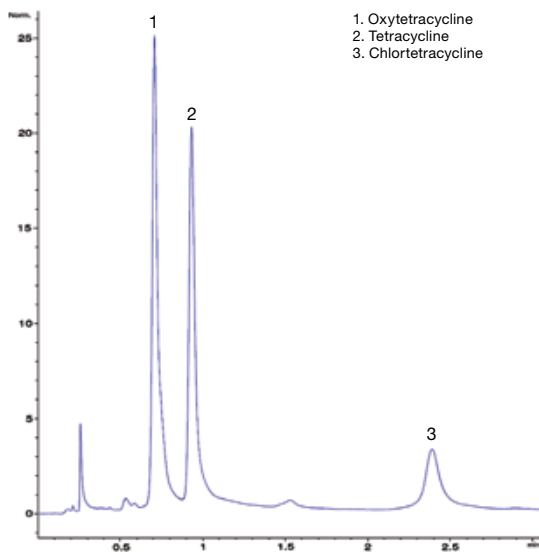
Sulpha drugs



Column: YMC-Triart C18 (12 nm, 1.9 μ m) 50 x 2.0 mm ID
Part No.: TA12SP9-0502PT
Eluent: H_2O + formic acid (pH 2.5) / acetonitrile (75/25)
Flow rate: 0.75 mL/min
Detection: UV at 280 nm
Pressure: 740 bar
Injection: 0.5 μ L
Temperature: 50 $^\circ\text{C}$

UHPLC

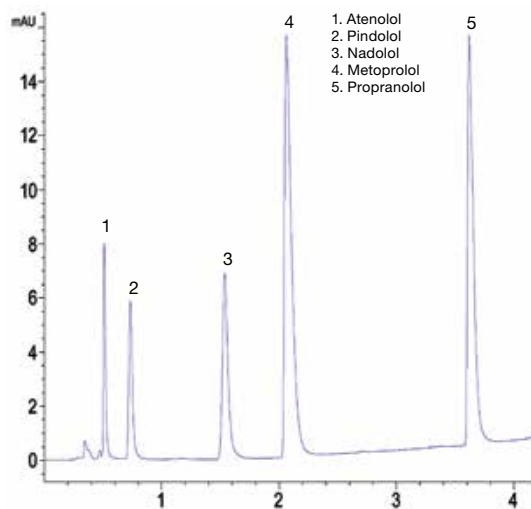
Tetracycline antibiotics



1. Oxytetracycline
2. Tetracycline
3. Chlortetracycline

Column: YMC-Triart C18 (12 nm, 1.9 μ m) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: 5 mM CH₃COONH₄ / acetonitrile (87/13)
 Flow rate: 0.65 mL/min
 Detection: UV at 280 nm
 Pressure: 662 bar
 Injection: 1 μ L
 Temperature: 40 °C

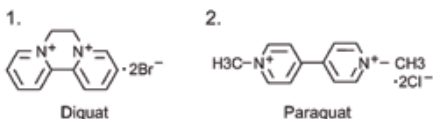
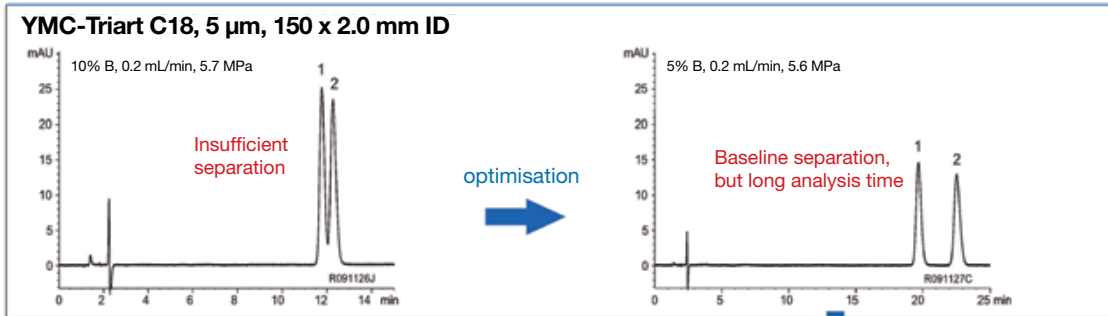
Betablockers



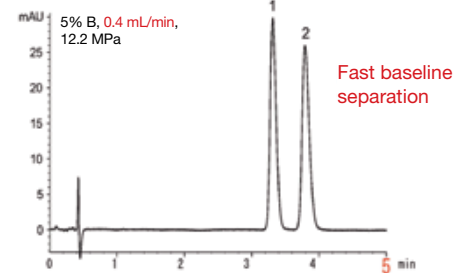
1. Atenolol
2. Pindolol
3. Nadolol
4. Metoprolol
5. Propranolol

Column: YMC-Triart C18 (12 nm, 1.9 μ m) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: A) 20 mM CH₃COONH₄ + ammonia (pH 9.0)
 B) acetonitrile
 Gradient: 25% B (1.0 min); 75% B (1-6 min)
 Flow rate: 0.35 mL/min
 Detection: UV at 254 nm
 Pressure: 450 bar
 Injection: 1 μ L
 Temperature: 40 °C

Fast LC for conventional HPLC*

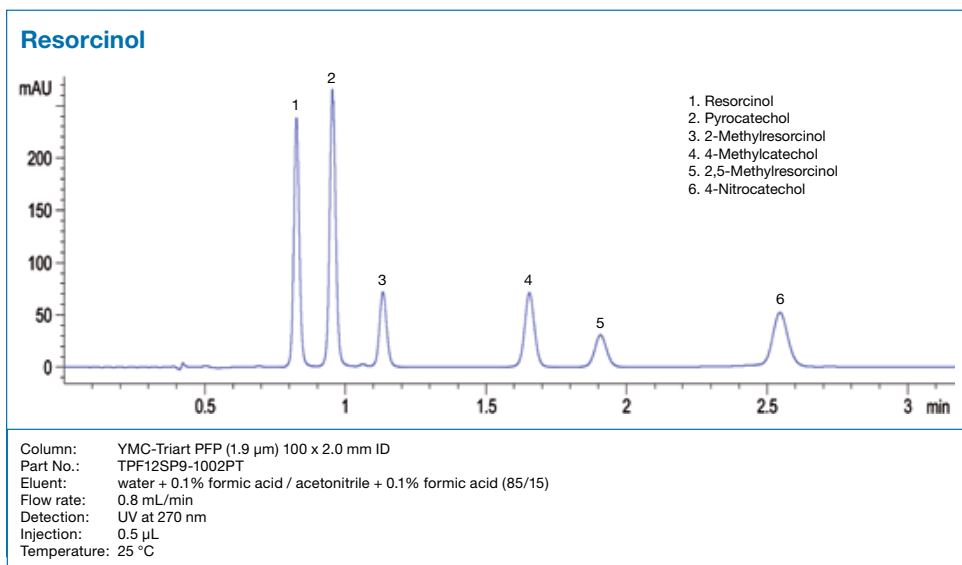
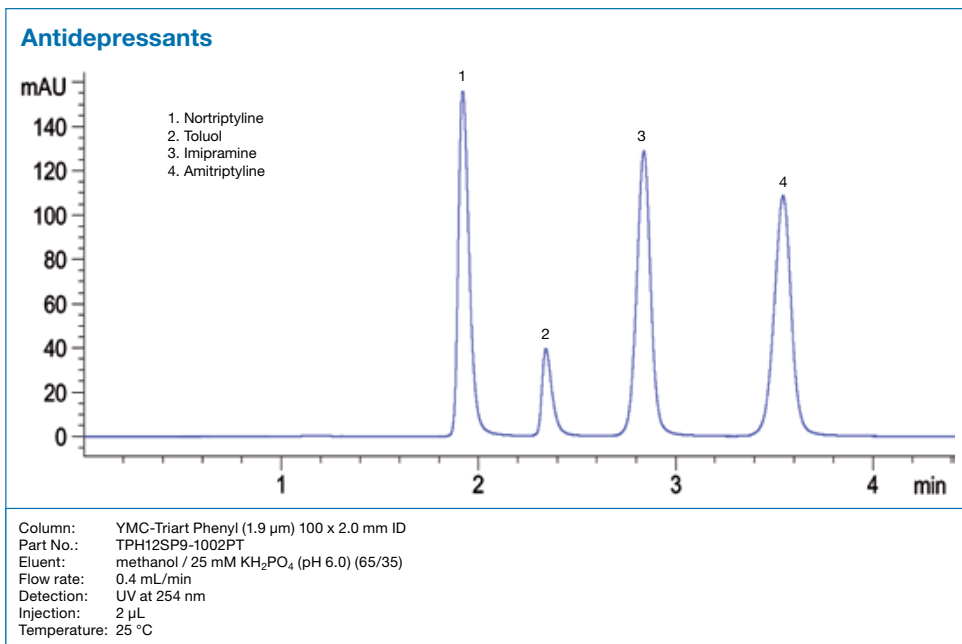


YMC-Triart C18, 3 μ m, 50 x 2.0 mm ID



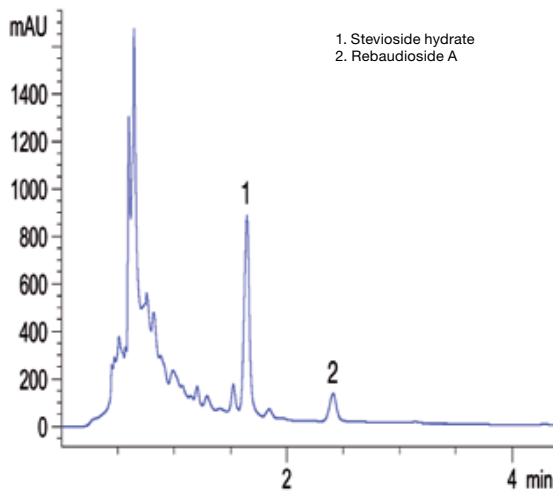
Eluent: A) water / HFBA* (100/0.1)
 B) acetonitrile / HFBA* (100/0.1)
 Temperature: 37 °C
 Detection: UV at 290 nm
 Injection: 1 μ L (0.1 mg/mL)
 *heptafluorobutyric acid

UHPLC



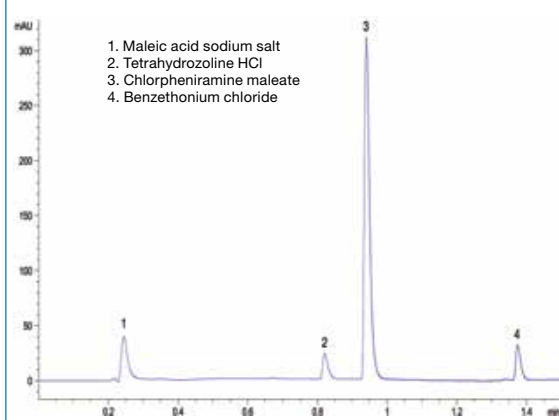
UHPLC

Stevia leaves



Column: YMC-Triart Diol-HILIC (1.9 μ m, 12 nm) 100 x 3.0 mm ID
 Part No.: TDH12SP9-1003PT
 Eluent: acetonitrile / water (85/15)
 Flow rate: 1 mL/min
 Detection: UV at 200 nm
 Injection: 2 μ L
 Temperature: 30 $^{\circ}$ C

Nasal spray

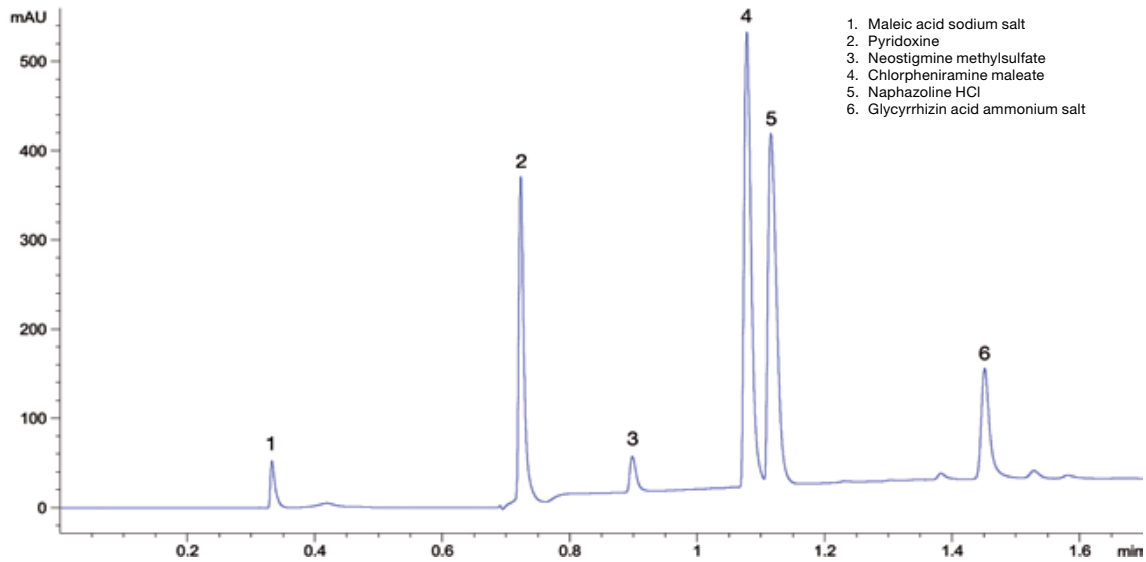


Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: A) water + 0.05% TFA / B) methanol (50/50)
 Gradient:

min	A	B
0	80	20
0.5	10	90
1.2	0	100

 Flow rate: 0.6 mL/min
 Detection: UV at 260 nm
 Injection: 0.2 μ L
 Temperature: 40 $^{\circ}$ C

Eye drop formulation



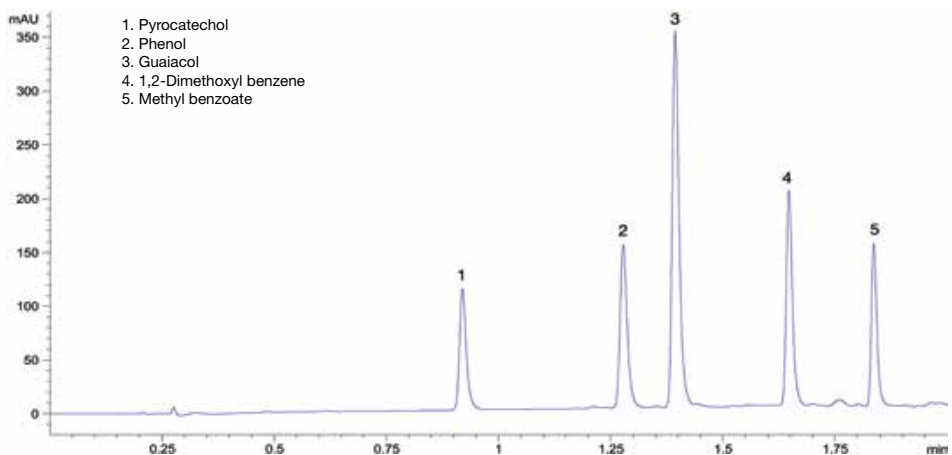
Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: A) water + 0.05% TFA
 B) acetonitrile
 Flow rate: 0.6 mL/min
 Detection: UV at 265 nm
 Injection: 0.5 μ L
 Temperature: 40 $^{\circ}$ C

Gradient:

min	A	B
0	100	0
1	50	50
1.5	50	50
1.7	10	90

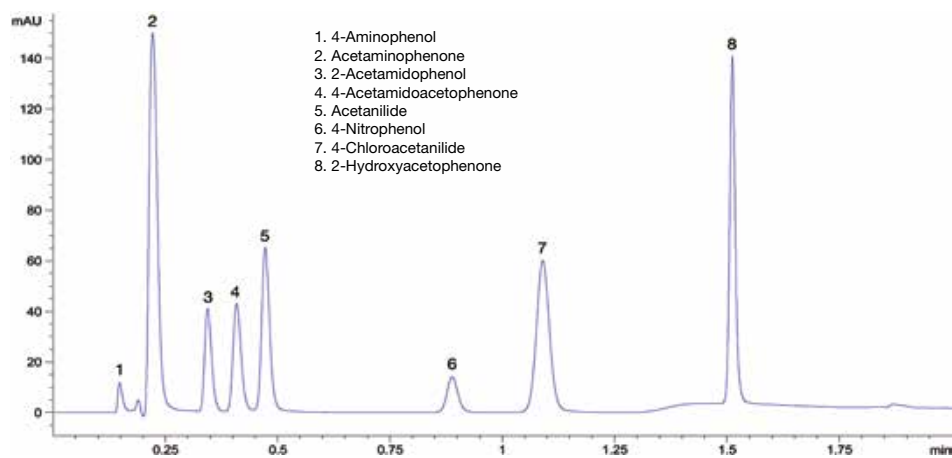
UHPLC

Guaiacol and impurities



Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: water / acetonitrile (50/50)
 Flow rate: 0.7 mL/min
 Detection: UV at 254 nm
 Injection: 0.5 μ L
 Temperature: 40 $^{\circ}$ C

Paracetamol

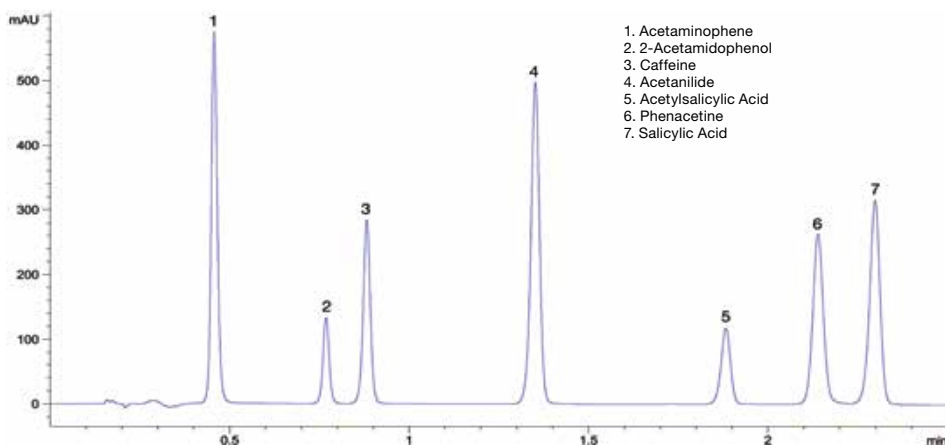


Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: A) water + formic acid (pH 2.5) / B) acetonitrile
 Flow rate: 0.7 mL/min
 Detection: UV at 254 nm
 Injection: 0.5 μ L
 Temperature: 40 $^{\circ}$ C

Gradient:	min	A	B
	0	70	30
	1	70	30
	1.5	20	80
	2	20	80

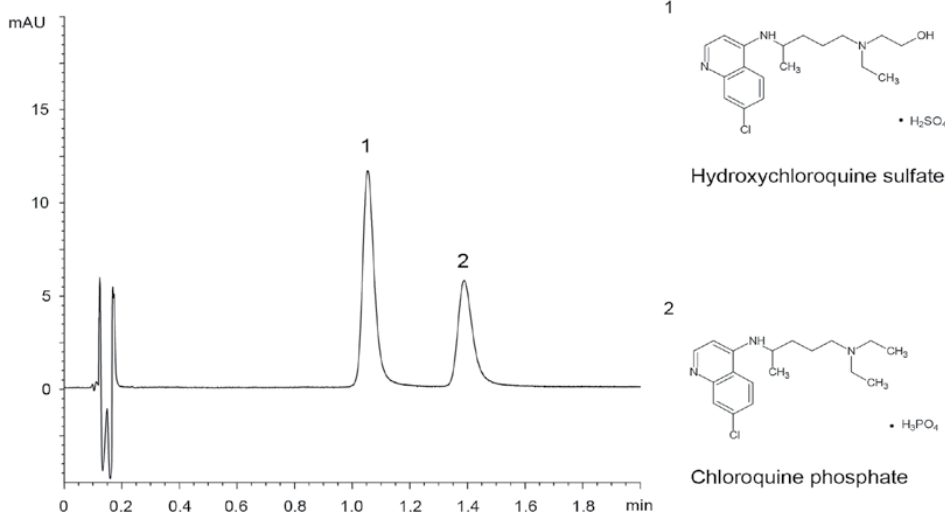
UHPLC

7 Analgesics



Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: water + formic acid (pH 2.5) / acetonitrile (50/50)
 Flow rate: 0.8 mL/min
 Detection: UV at 240 nm
 Injection: 1 μ L
 Temperature: 40 $^{\circ}$ C

Hydroxychloroquine and chloroquine

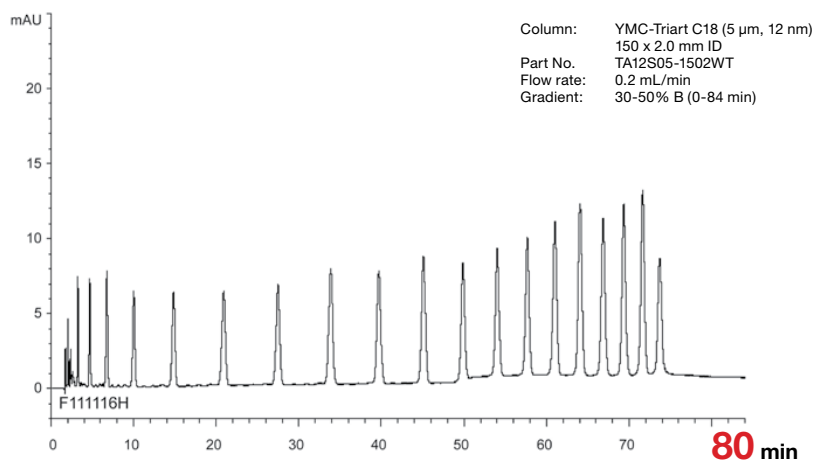


Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: 20 mM HCOOH-HCOONH₄ (pH 4.3) / acetonitrile (90/10)
 Flow rate: 1.0 mL/min
 Detection: UV at 254 nm
 Injection: 2 μ L (10 μ g/mL)
 Temperature: 25 $^{\circ}$ C

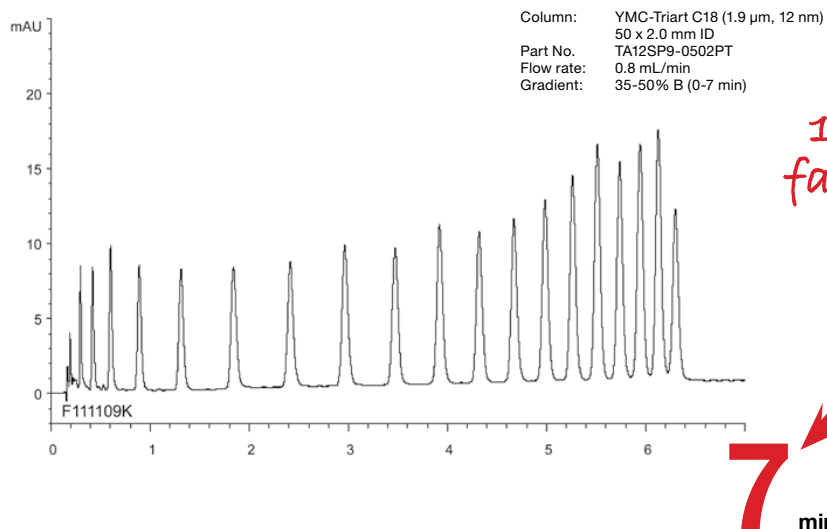
UHPLC

Oligonucleotides d(T)₂₋₂₀ method transfer from HPLC to UHPLC*

Conventional LC method



UHPLC method

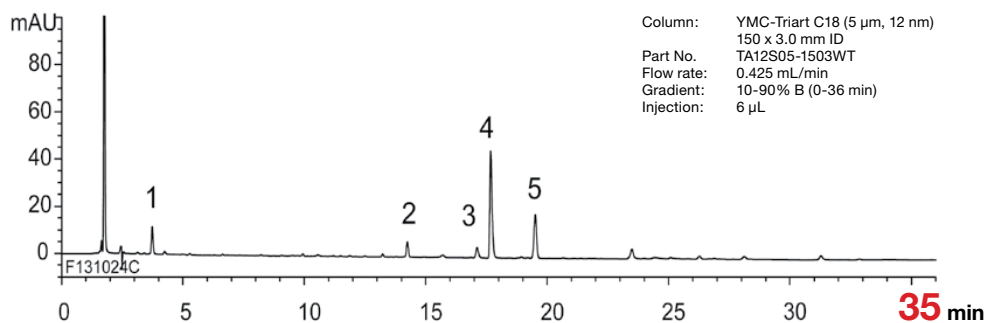


Eluent: A) 10 mM di-n-butylamine-acetic acid (pH 6.0)
B) methanol
Detection: UV at 269 nm
Injection: 1 μ L (5 nmol/mL)
Temperature: 37 °C

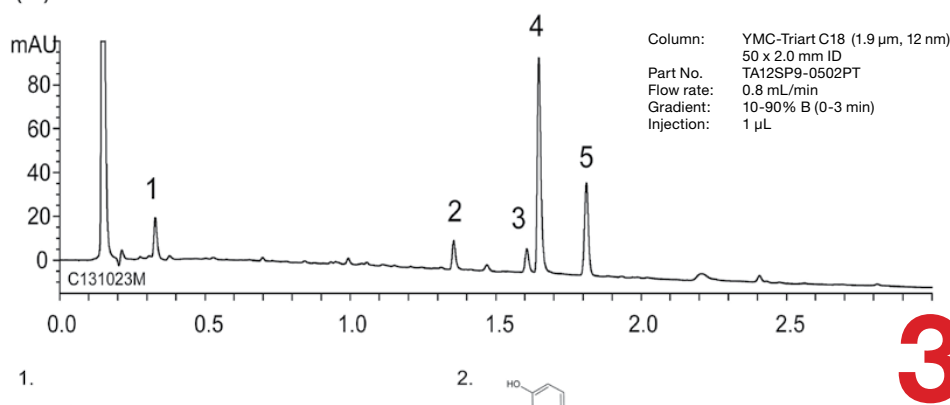
UHPLC

Duloxetine and its degradation products

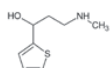
(A) HPLC method



(B) UHPLC method

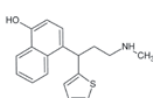


1.



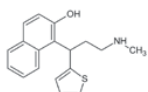
Amino alcohol
 (3-Methylamino-1-thiophen-2-yl-propan-1-ol)

2.



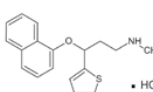
Para isomer
 (4-(3-Methylamino-1-thiophen-2-yl-propyl)-naphthalen-1-ol)

3.



Ortho isomer
 (2-(3-Methylamino-1-thiophen-2-yl-propyl)-naphthalen-1-ol)

4.



Duloxetine hydrochloride

5.

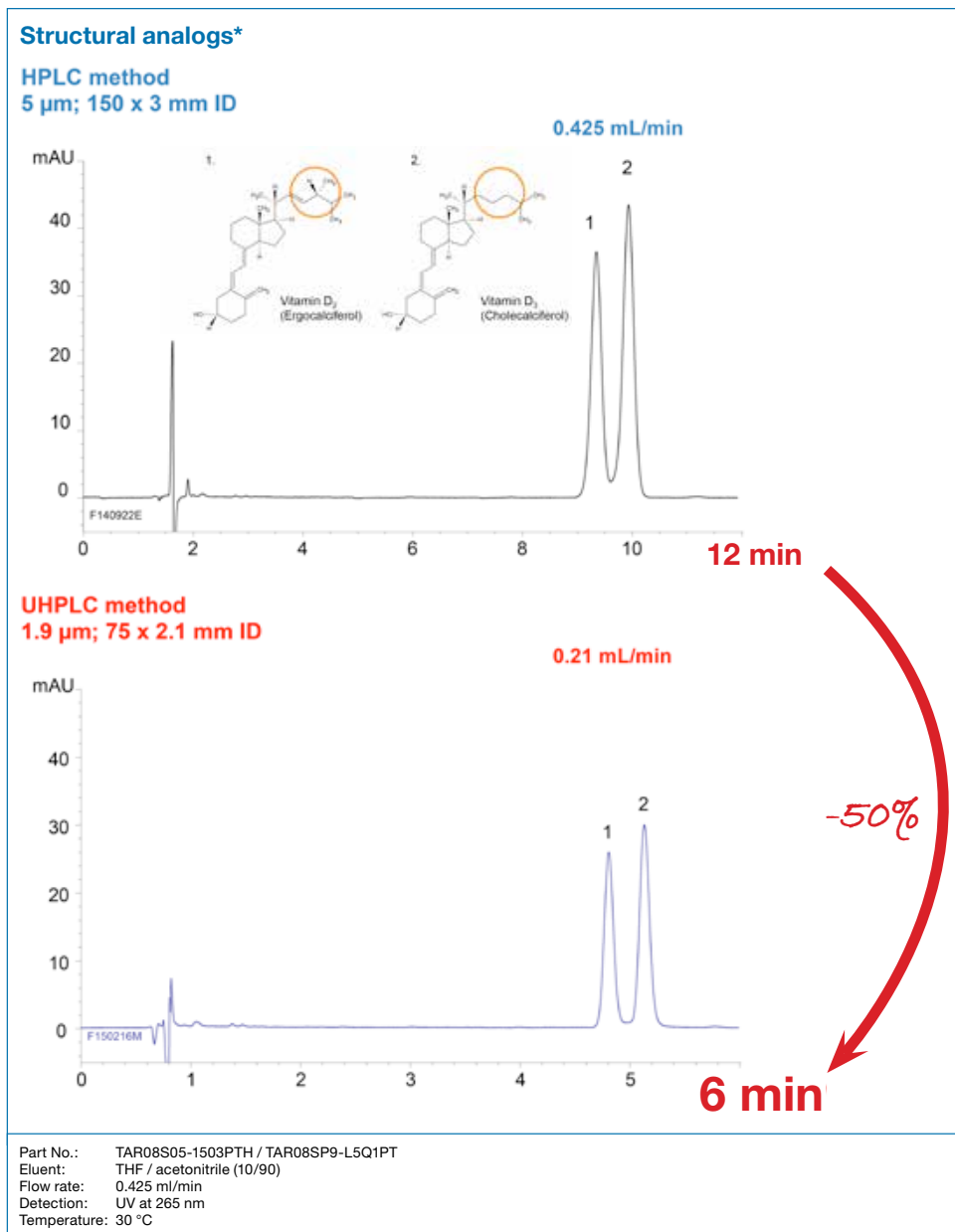


α-Naphthol

Eluent: A) 10 mM CH₃COONH₄ (pH 6.0)
 B) acetonitrile
 Detection: UV at 230 nm
 Temperature: 30 °C
 Sample: Oxidative degradation products of duloxetine hydrochloride*

* Sample preparation was performed as described by Veera Reddy. Arava et al. Der Pharma Chemica, 2012 4 (4): 1735-1741

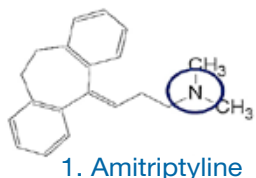
YMC-Triart C18 ExRS



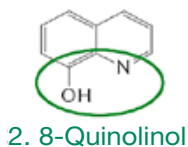
YMC-Triart C18 ExRS

High hydrophobicity & high steric recognition ability*

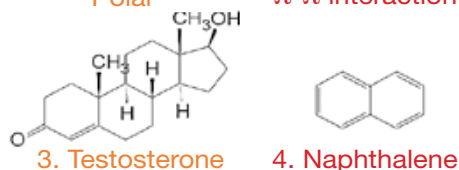
Basic Compound



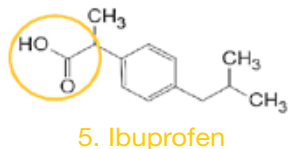
Coordination Compound



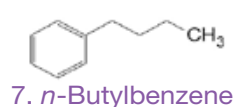
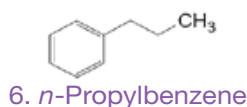
Neutral Compounds
Polar π - π interaction



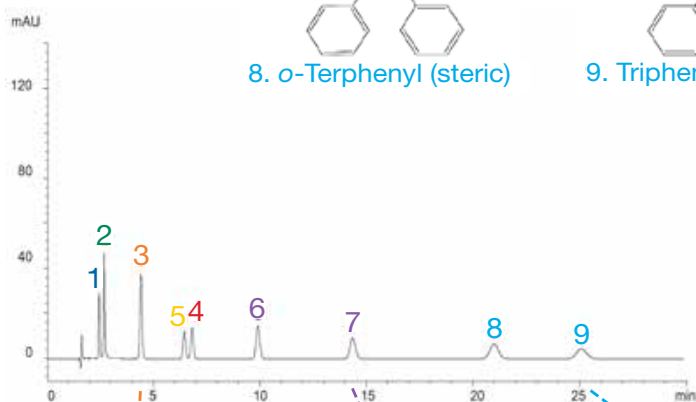
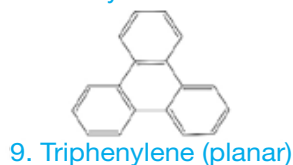
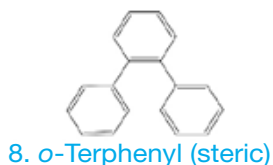
Acidic Compound



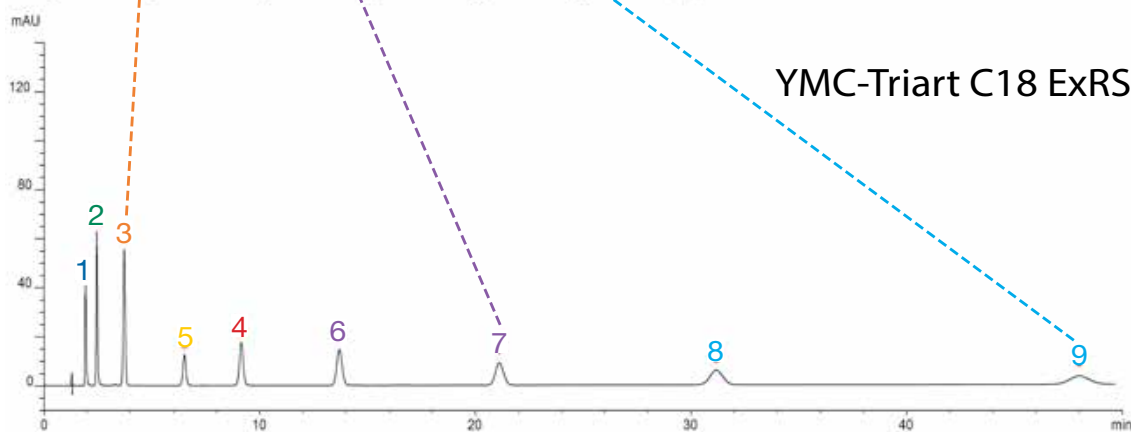
Hydrophobic



Steric Cognitive Ability



YMC-Triart C18



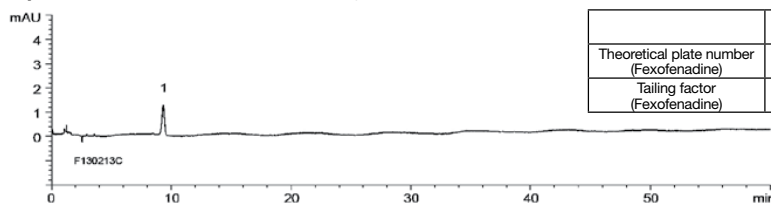
YMC-Triart C18 ExRS

Column: 5 μ m, 150 x 3.0 ID
 Part No.: TA12S05-1503PTH / TAR08S05-1503PTH
 Eluent: 20 mM HCOOH-HCOONH₄ (pH 4.3) / acetonitrile (90/10)
 Flow rate: 1.0 mL/min
 Detection: UV at 254 nm
 Injection: 2 μ L (10 μ g/mL)
 Temperature: 25 °C

YMC-Triart Phenyl

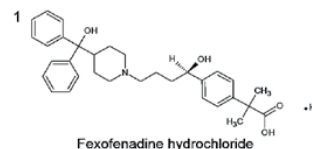
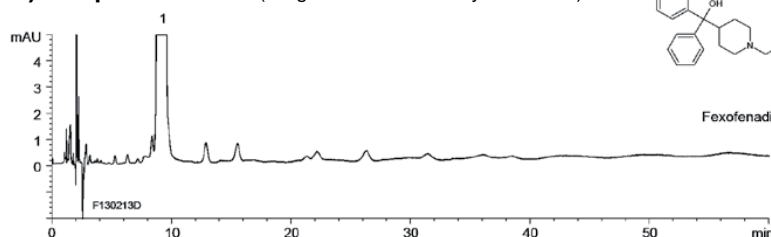
Fexofenadine hydrochloride (Japanese Pharmacopoeia)*

A) Standard solution *1 (0.001 mg/mL fexofenadine hydrochloride)



	System suitability requirement	result
Theoretical plate number (Fexofenadine)	≥ 8000	10100
Tailing factor (Fexofenadine)	≤ 2.0	1.00

B) Sample solution *1 (1 mg/mL fexofenadine hydrochloride)

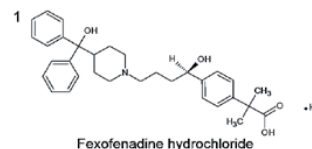
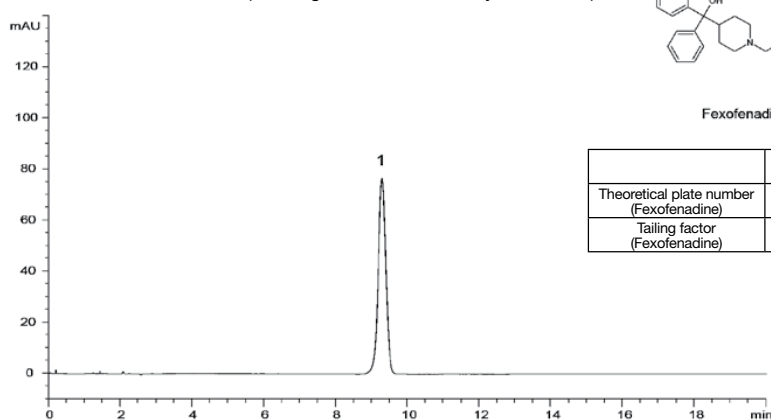


Column: YMC-Triart Phenyl (5 μm, 12 nm) 250 x 4.6 mm ID
 Part No.: TPH12S05-2546WT
 Eluent: acetonitrile / buffer *2 / triethylamine (350/650/3)
 *2 Dissolve 7.51 g of NaH₂PO₄·2H₂O and 0.96 g of NaClO₄·H₂O in 1000 mL water, adjust pH 2.0 with H₃PO₄
 Flow rate: 2.0 mL/min (adjust the flow rate so that the retention time of fexofenadine is about 9 min)
 Detection: UV at 220 nm
 Injection: 20 μL
 Temperature: 25 °C
 (The Japanese Pharmacopoeia 16th; related substances)

*1 All standard and sample solutions were prepared from fexofenadine hydrochloride supplied as a reagent for laboratory use.

Fexofenadine hydrochloride (Japanese Pharmacopoeia)*

Standard solution *1 (0.06 mg/mL fexofenadine hydrochloride)



	System suitability requirement	result
Theoretical plate number (Fexofenadine)	≥ 8000	9500
Tailing factor (Fexofenadine)	≤ 2.0	0.98

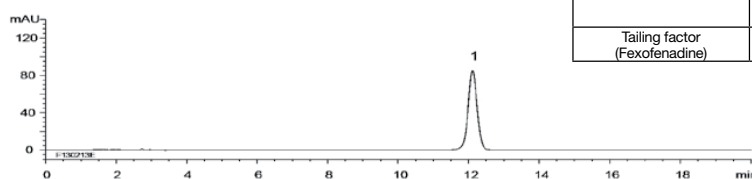
Column: YMC-Triart Phenyl (5 μm, 12 nm) 250 x 4.6 mm ID
 Part No.: TPH12S05-2546WT
 Eluent: acetonitrile / buffer *2 / triethylamine (350/650/3)
 *2 Dissolve 7.51 g of NaH₂PO₄·2H₂O and 0.96 g of NaClO₄·H₂O in 1000 mL water, adjust pH 2.0 with H₃PO₄
 Flow rate: 2.0 mL/min (adjust the flow rate so that the retention time of fexofenadine is about 9 min)
 Detection: UV at 220 nm
 Injection: 20 μL
 Temperature: 25 °C
 (The Japanese Pharmacopoeia 16th; assay)

*1 Standard solutions was prepared from fexofenadine hydrochloride supplied as a reagent for laboratory use.

YMC-Triart Phenyl

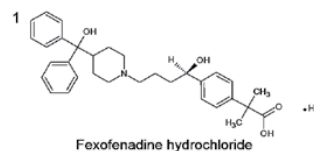
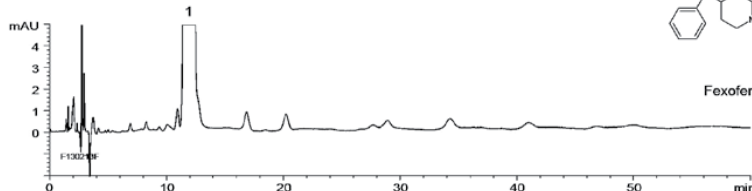
Fexofenadine hydrochloride (US Pharmacopoeia)*

A) Assay preparation *1 (assay), **Reference solution *1** (related compounds)
(0.06 mg/mL fexofenadine hydrochloride)



	System suitability requirement (assay)	result
Tailing factor (Fexofenadine)	≤ 2.0	1.00

B) Test solution *1 (related compounds)
(1 mg/mL fexofenadine hydrochloride)

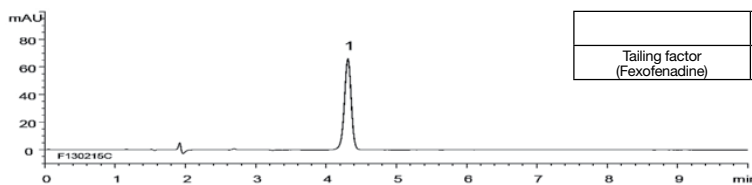


Column: YMC-Triart Phenyl (5 μ m, 12 nm) 250 x 4.6 mm ID
Part No.: TPH12S05-2546WT
Eluent: acetonitrile / buffer *2 / triethylamine (350/650/3)
*2 Dissolve 7.51 g of NaH₂PO₄·2H₂O and 0.96 g of NaClO₄·H₂O in 1000 mL water, adjust pH 2.0 with H₃PO₄
Flow rate: 1.5 mL/min
Detection: UV at 220 nm
Injection: 20 μ L
Temperature: 25 °C
(The United States Pharmacopoeia 36th; assay, related compounds)

*1 All standard and sample solutions were prepared from fexofenadine hydrochloride supplied as a reagent for laboratory use.

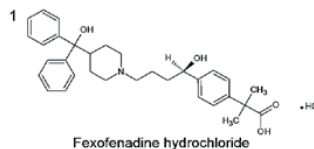
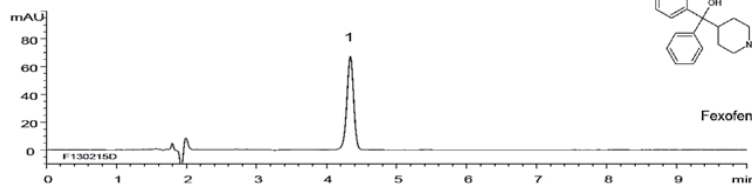
Fexofenadine hydrochloride (US Pharmacopoeia)*

A) Standard solution *1 (0.015 mg/mL fexofenadine hydrochloride)



	System suitability requirement (assay)	result
Tailing factor (Fexofenadine)	≤ 2.0	0.95

B) Sample solution *2 (0.018 mg/mL fexofenadine hydrochloride)



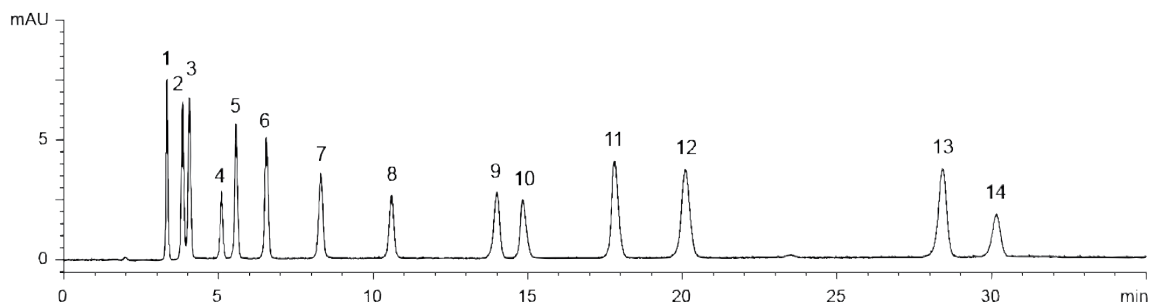
Column: YMC-Triart Phenyl (5 μ m, 12 nm) 250 x 4.6 mm ID
Part No.: TPH12S05-2546WT
Eluent: acetonitrile / buffer *3 (9/16)
*3 Add 15 mL of acetonitrile/triethylamine (1/1) to 1000 mL of acetic acid/water (17/9983), adjust pH 5.25 with H₃PO₄
Flow rate: 1.5 mL/min
Detection: UV at 220 nm
Injection: 20 μ L
Temperature: 35 °C
(The United States Pharmacopoeia 36th; assay)

*1 Standard solution was prepared from fexofenadine hydrochloride supplied as a reagent for laboratory use.

*2 Sample solution was prepared from fexofenadine hydrochloride tablets.

YMC-Triart PFP

Catecholamines, serotonin, and their precursors and metabolites*

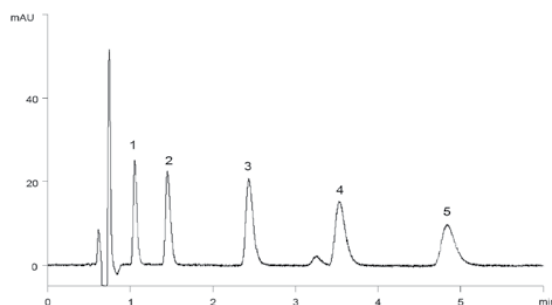


<p>1</p> <p>Noradrenaline hydrochloride (NA) (Norepinephrine hydrochloride)</p>	<p>2</p> <p>3,4-Dihydroxymandelic acid (DOMA)</p>	<p>3</p> <p>3,4-Dihydroxyphenylalanine (DOPA)</p>	<p>4</p> <p>Tyrosine (Tyr)</p>
<p>5</p> <p>Adrenaline hydrochloride (A) (Epinephrine hydrochloride)</p>	<p>6</p> <p>Dopamine hydrochloride (DA)</p>	<p>7</p> <p>Vanillylmandelic acid (VMA)</p>	<p>8</p> <p>3-Methoxy-4-hydroxyphenylglycol (MHPG)</p>
<p>9</p> <p>3,4-Dihydroxyphenylacetic acid (DOPAC)</p>	<p>10</p> <p>3-Methoxytyramine hydrochloride (3MT)</p>	<p>11</p> <p>Serotonin hydrochloride (5-Hydroxytryptamine hydrochloride, 5HT)</p>	<p>12</p> <p>Tryptophan (Trp)</p>
<p>13</p> <p>5-Hydroxyindoleacetic acid (5HIAA)</p>	<p>14</p> <p>Homovanillic acid (HVA)</p>		

Column: YMC-Triart PFP (3 μ m, 12 nm) 150 x 3.0 mm ID
 Part No.: TPF12S03-1503WT
 Eluent: A) 10 mM formic acid
 B) methanol containing 10 mM formic acid
 0-20% B (0-30 min), 20% B (30-35 min)
 Flow rate: 0.425 mL/min
 Detection: UV at 280 nm
 Injection: 4 μ L (5 μ g/mL)
 Temperature: 25 $^{\circ}$ C

Pharmaceuticals

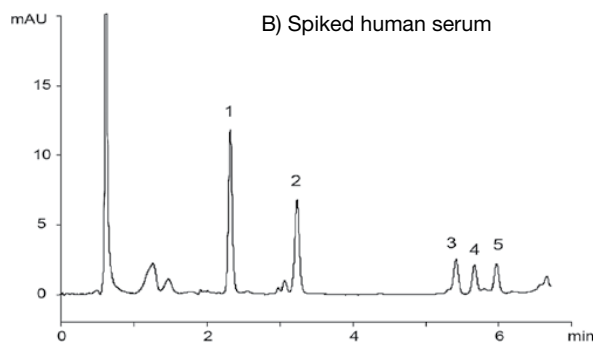
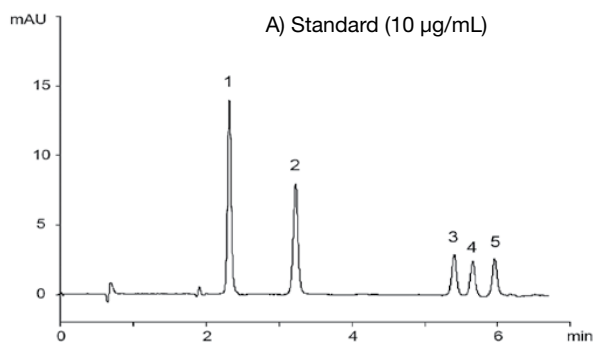
Separation of alkaloids*



1. Scopolamine
2. Atropine
3. Cinchonine
4. Quinine
5. Dihydroquinine

Column: YMC-Triart C18 (5 μ m, 12 nm)
 50 x 2.0 mm ID
 Part No.: TA12S05-0502WT
 Eluent: 20 mM CH₃COOH-CH₃COONH₄
 (pH 4.9) / acetonitrile (80/20)
 Flow rate: 0.2 mL/min
 Temperature: 40 °C
 Detection: UV at 220 nm
 Injection: 1 μ L (0.02-0.1 mg/mL)

Barbiturates in human serum*



Solid-phase extraction method

YMC Dispo SPE C18 100 mg/1mL

Condition

2 mL methanol
 2 mL water

Load

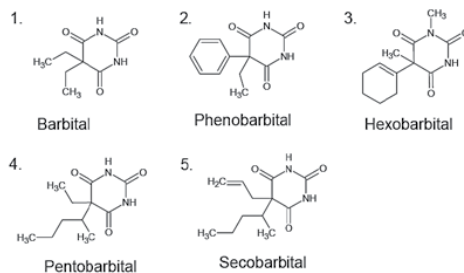
500 μ L spiked human serum
 solution (each 10 μ g)

Elute

500 μ L methanol/water (85/15)

Dilute

500 μ L 20 mM ammonium
 formate buffer (pH 9.5)

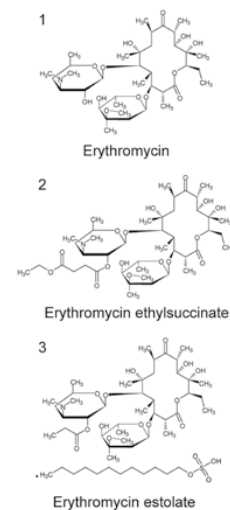
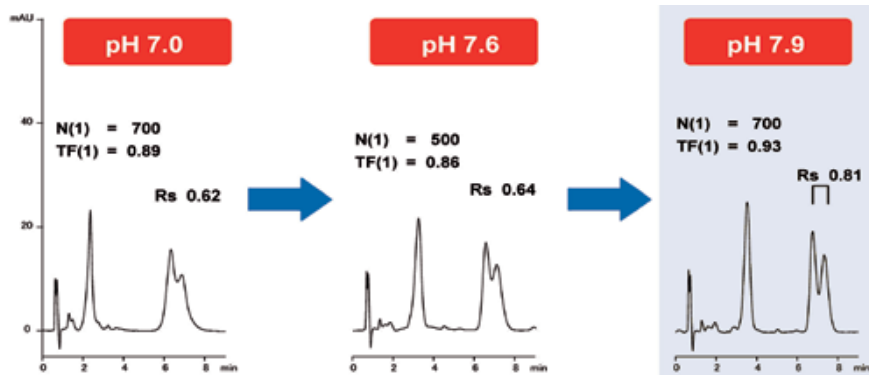


Column: YMC-Triart C18 (5 μ m, 12 nm)
 50 x 2.0 mm ID
 Part No.: TA12S05-0502WT
 Eluent: A) 20 mM HCOONH₄-NH₃ (pH 9.5)
 B) methanol
 Gradient: 0-90% B (0-7 min)
 Flow rate: 0.2 mL/min
 Temperature: 25 °C
 Detection: UV at 240 nm
 Injection: 1 μ L

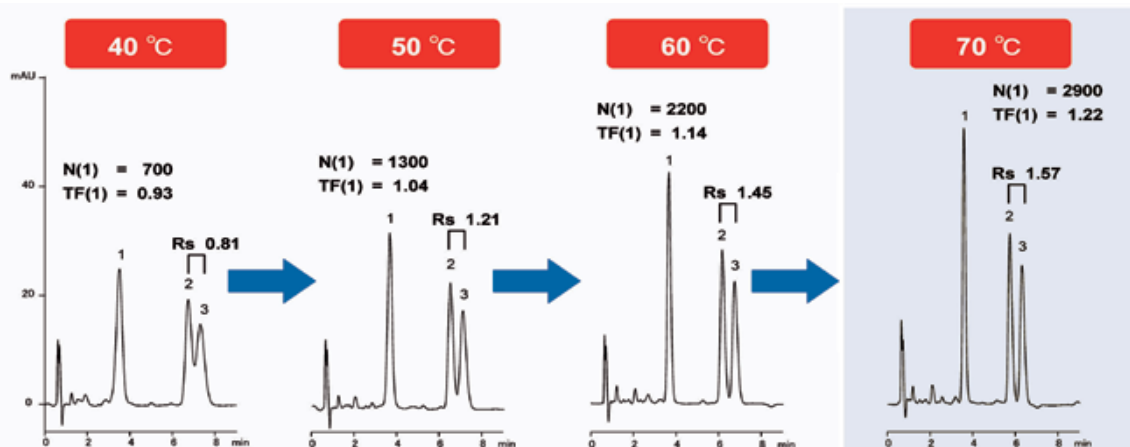
Pharmaceuticals

Erythromycin at elevated pH and temperature*

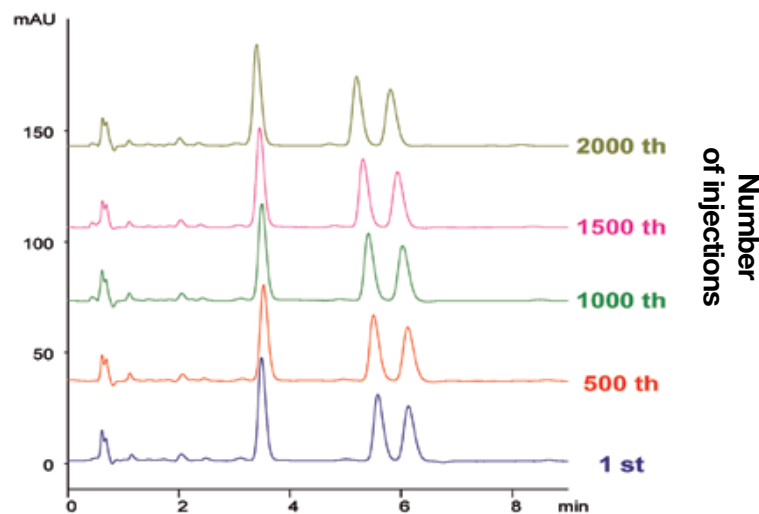
1. Optimisation of pH



2. Optimisation of temperature (pH 7.9)



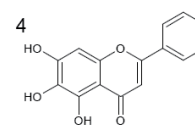
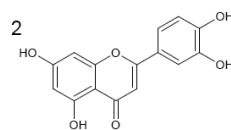
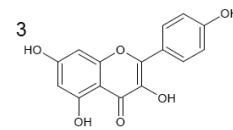
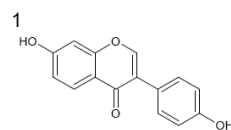
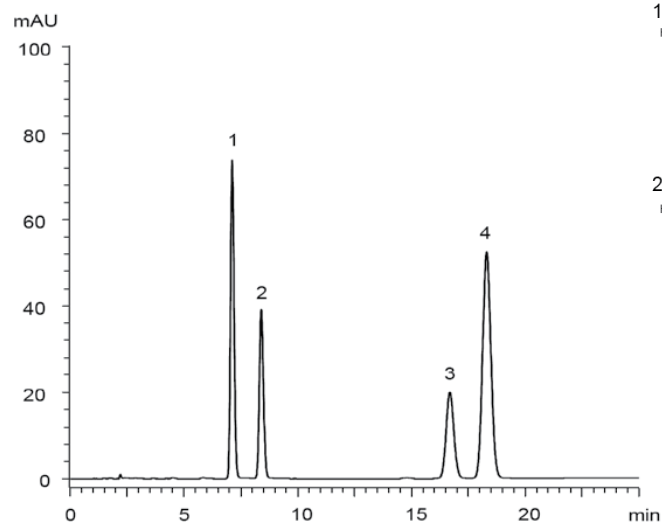
3. Stability test: pH 7.9, 70 °C



Column: YMC-Triart C18 (3 μ m, 12 mm)
50 x 2.0 mm ID
Part No.: TA12S03-0502WT
Eluent: 20 mM KH_2PO_4 - K_2HPO_4 / acetonitrile / methanol (40/45/15)
Flow rate: 0.2 mL/min
Detection: UV at 210 nm

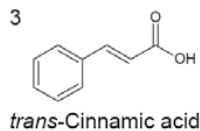
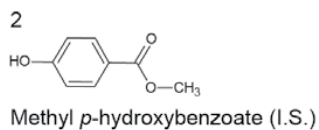
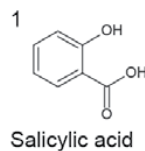
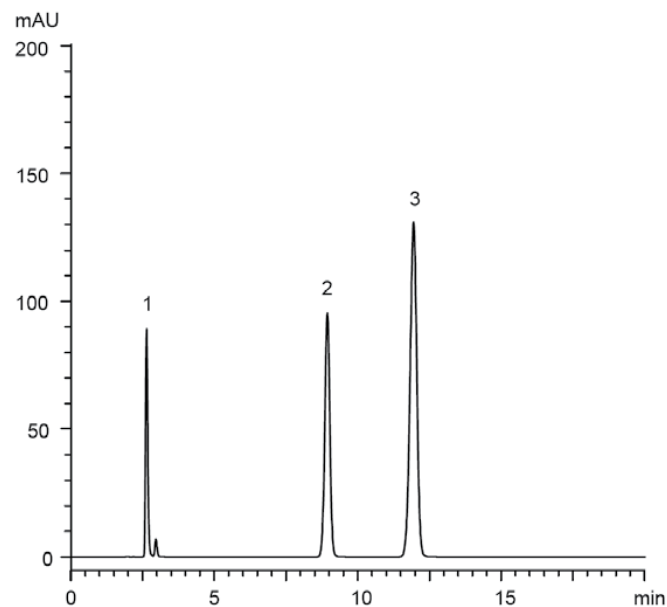
Pharmaceuticals

Separation of flavonoids*



Column: YMC-Triart C18 (5 μ m, 12 nm)
 150 x 3.0 mm ID
 Part No.: TA12S05-1503WT
 Eluent: acetonitrile / 10 mM H₃PO₄ (30/70)
 Flow rate: 0.425 mL/min
 Temperature: 37 °C
 Detection: UV at 280 nm
 Injection: 2 μ L (50 μ g/mL)

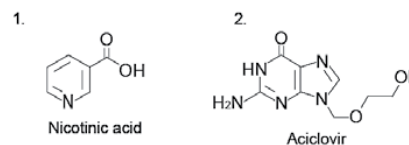
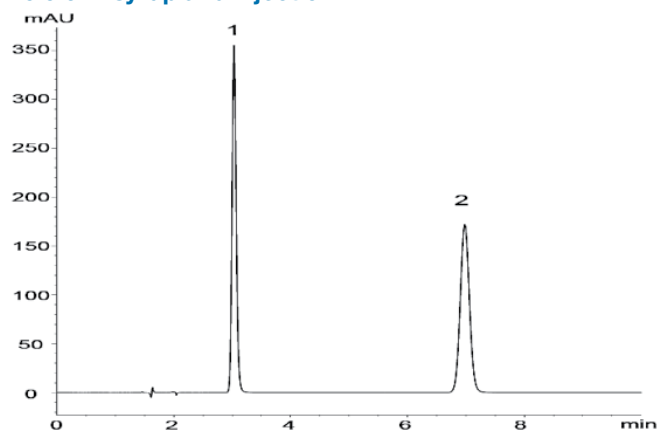
Separation of aromatic carboxylic acids*



Column: YMC-Triart C18 (5 μ m, 12 nm)
 150 x 3.0 mm ID
 Part No.: TA12S05-1503WT
 Eluent: 10 mM CH₃COOH-CH₃COONH₄ (pH 4.2) /
 acetonitrile (75/25)
 Flow rate: 0.425 mL/min
 Temperature: 40 °C
 Detection: UV at 254 nm
 Injection: 4 μ L (0.02 - 0.3 mg/mL)

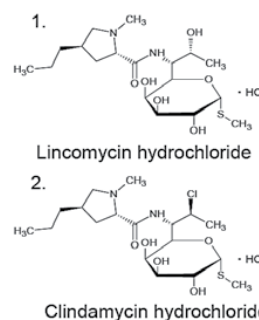
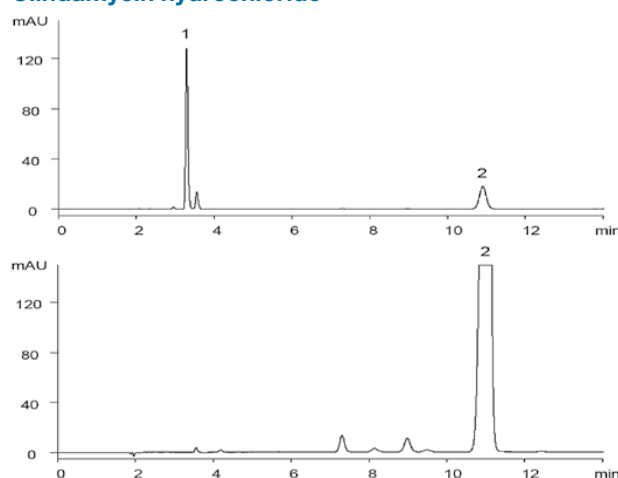
Pharmaceuticals

Aciclovir syrup and injection*



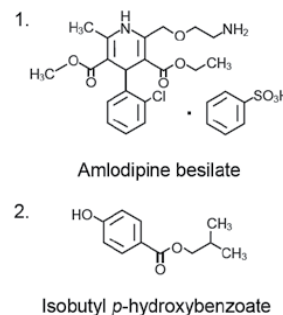
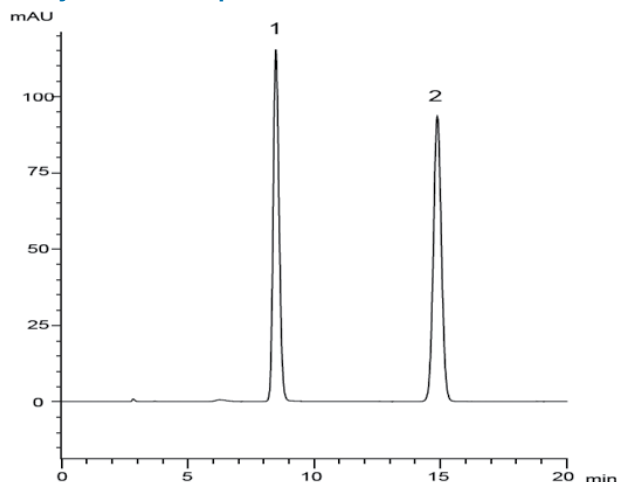
Column: YMC-Triart C18 (5 μ m, 12 nm) 150 x 4.6 mm ID
 Part No.: TA12S05-1546WT
 Eluent: phosphate buffer* / methanol (95/5)
 *Dissolve 1.45 g of H₃PO₄ and 25 mL of 1 mol/l CH₃COOH in water to make 900 mL \rightarrow adjust pH 2.5 by 1 mol/l NaOH \rightarrow add water to make 1000 mL
 Flow rate: 1.0 mL/min
 Temperature: 25 $^{\circ}$ C
 Detection: UV at 254 nm
 Injection: 20 μ L (0.05 mg/mL, 0.032 mg/mL)

Clindamycin hydrochloride*



Column: YMC-Triart C18 (5 μ m, 12 nm) 250 x 4.6 mm ID
 Part No.: TA12S05-2546WT
 Eluent: 50 mM KH₂PO₄ (pH 7.5 adjusted by 8 M KOH) / acetonitrile (55/45)
 Flow rate: 1.0 mL/min
 Temperature: 25 $^{\circ}$ C
 Detection: UV at 210 nm
 Injection: 10 μ L

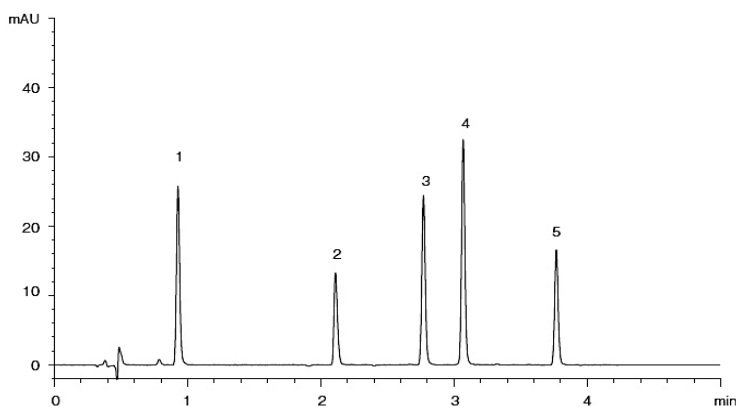
Analysis of amlodipine besilate*



Column: YMC-Triart C18 (5 μ m, 12 nm) 150 x 3.0 mm ID
 Part No.: TA12S05-1503WT
 Eluent: 10 mM CH₃COOH-CH₃COONH₄ (pH 4.2) / acetonitrile (75/25)
 Flow rate: 0.425 mL/min
 Temperature: 40 $^{\circ}$ C
 Detection: UV at 254 nm
 Injection: 4 μ L (0.02 – 0.3 mg/mL)

Pharmaceuticals

Basic drugs*



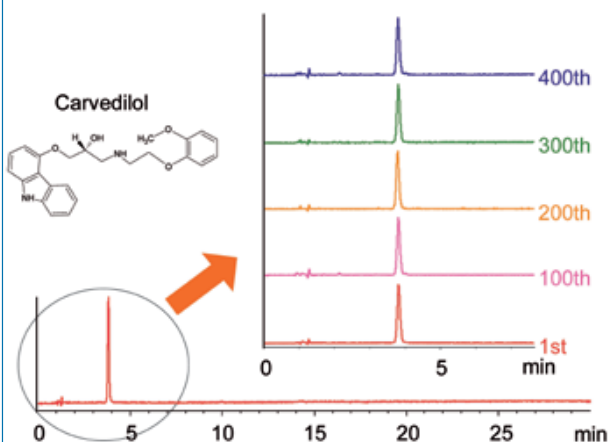
1. Hydrochlorothiazide
2. Amlodipine besilate
3. Valsartan
4. Atorvastatin calcium hydrate
5. Candesartan cilexetil

YMC-Triart C8

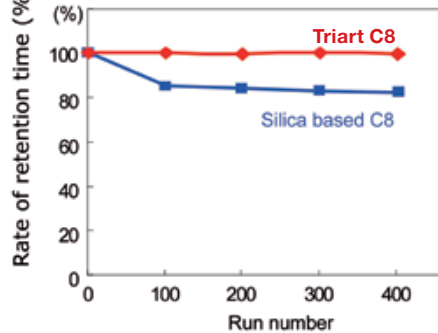
Column: YMC-Triart C8 (3 μ m, 12 nm), 50 x 2.0 mm ID
 Part No.: TO12S03-0502WT
 Eluent: A) water / formic acid (100/0.1)
 B) acetonitrile / formic acid (100/0.1)
 10-90% B (0-5 min), 90% B (5-7 min)

Flow rate: 0.4 mL/min
 Temperature: 30 °C
 Detection: UV at 254 nm
 Injection: 2 μ L (10-20 μ g/mL)

Sequential analysis of Carvedilol*



Retention stability of carvedilol

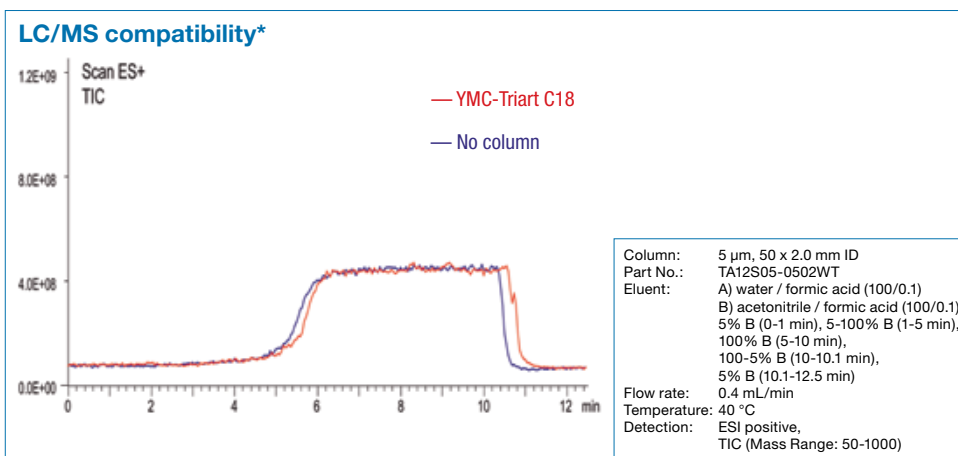


Column: YMC-Triart C8 (5 μ m, 150 x 2.0 mm ID)
 Part No.: TO12S05-1502WT
 Eluent: phosphate buffer (pH 2.0)* / acetonitrile (65/35)
 *Dissolve 2.72 g of KH_2PO_4 in 900 mL water, adjust pH 2.0 with H_3PO_4 and add water to make 1000 mL
 Flow rate: 0.28 mL/min (adjust the flow rate so that the retention time of carvedilol is about 4 min)
 Temperature: 55 °C
 Detection: UV at 240 nm

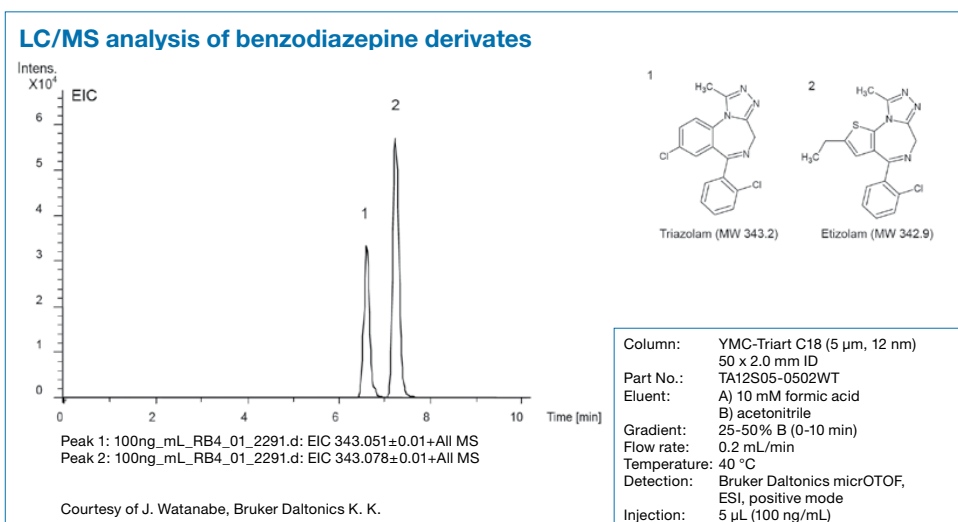
YMC-Triart C8

No change in retention time is observed even under a high pH and at an elevated temperature.

LC/MS



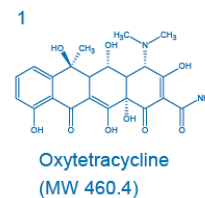
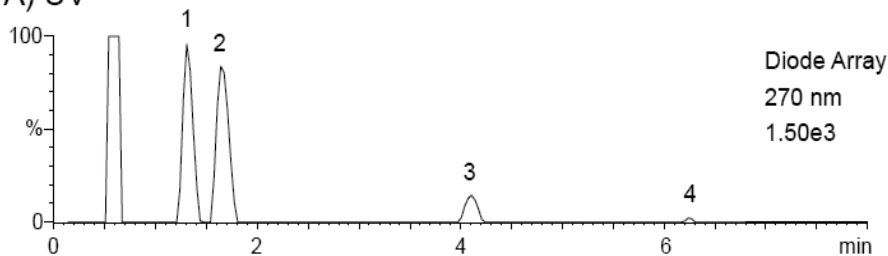
Column bleeding, caused by the fragments of stationary phase, is the main reason for background noise and restrictions on detection limits. No bleed is observed in the test of total ion current (TIC) measured by LC/MS with blank or with YMC-Triart C18. So in terms of the signal/noise ratio (S/N ratio), YMC-Triart C18 can be expected to not only reduce the background noise but to also increase the sensitivity of the analysis.



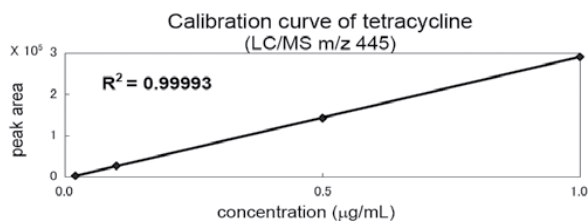
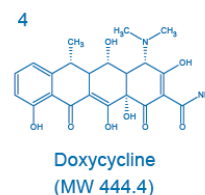
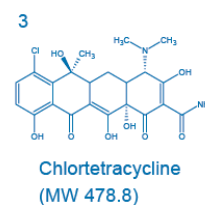
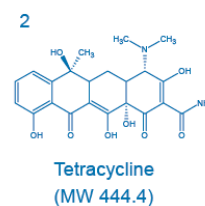
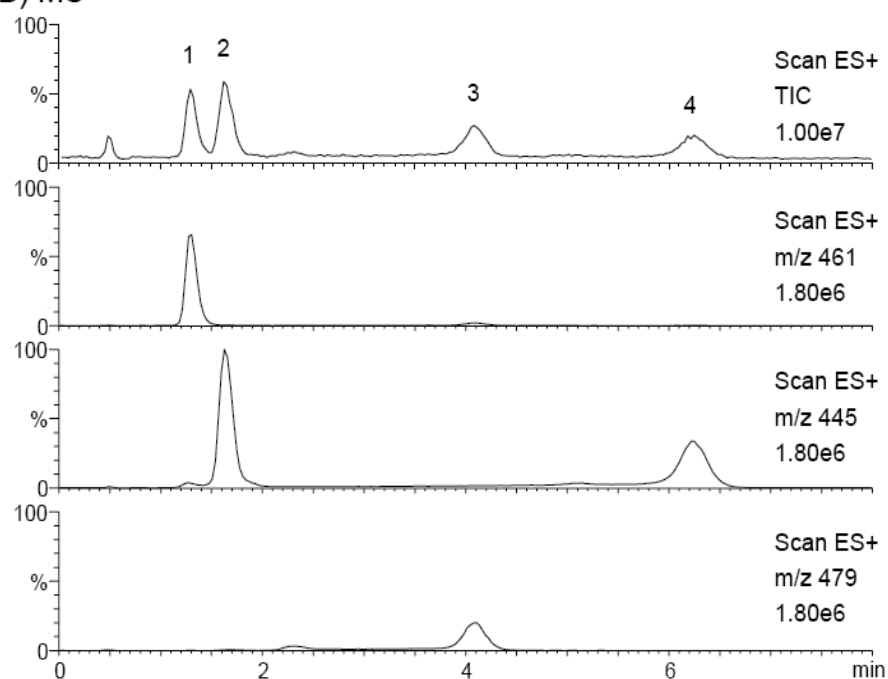
LC/MS

LC/MS analysis of tetracycline antibiotics*

A) UV



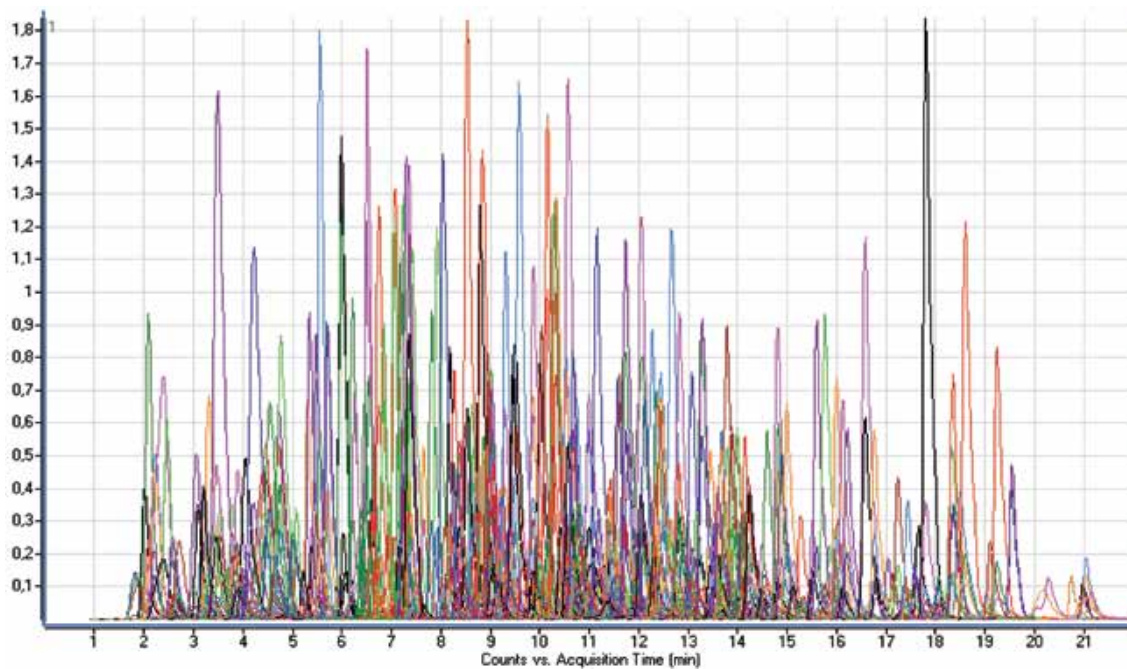
B) MS



Column: YMC-Triart C18 (5 μm , 12 nm) 50 x 2.0 mm ID
Part No.: TA12S05-0502WT
Eluent: acetonitrile / water / formic acid (15/85/0.1)
Flow rate: 0.4 mL/min
Temperature: 40 $^{\circ}\text{C}$
Detection: A) UV at 270 nm
B) ESI positive-mode
Injection: 10 μL (1 $\mu\text{g/mL}$)

LC/MS

Analysis of 360 pesticides in a single run

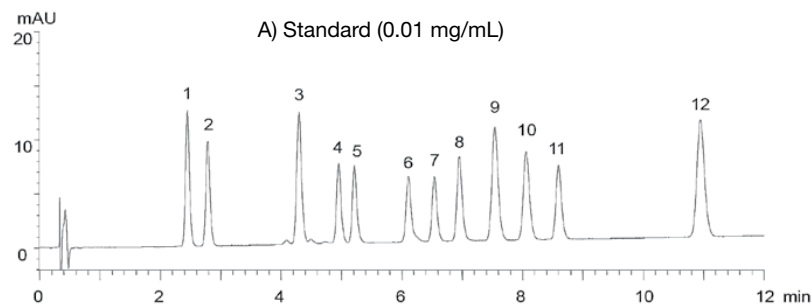


Column:	YMC-Triart C18 (3 µm) 100 x 2.0 mm ID	Injection:	5 µL
Part No.:	TA12S03-1002WT	Gradient:	0 min: 30% B, 0.1 min: 50% B, 18 min: 100% B, 21 min: 100% B, 21.01 min: 30% B, 29 min: 30% B
Eluent:	A) 5 mM ammonium formate / water B) 5 mM ammonium formate / methanol	Total run time:	30 min
Flow rate:	0.25 mL/min	Sample:	100 ng/mL pesticide mix in acetonitrile
Temperature:	45 °C		

by courtesy of: József László
WIREC, WESSLING International Research and Educational Centre Nonprofit Co. (Hungary)

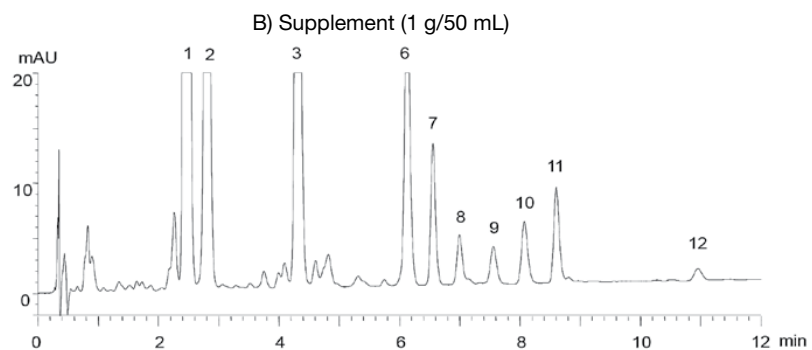
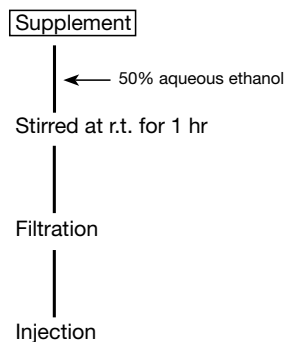
Food

Soy isoflavones in supplement*



1. Daidzin
2. Glycitin
3. Genistin
4. 6"-O-Malonyldaidzin
5. 6"-O-Malonylglycitin
6. 6"-O-Acetyldaidzin
7. 6"-O-Acetylglycitin
8. 6"-O-Malonylgenistin
9. Daidzein
10. Glycitein
11. 6"-O-Acetylgenistin
12. Genistein

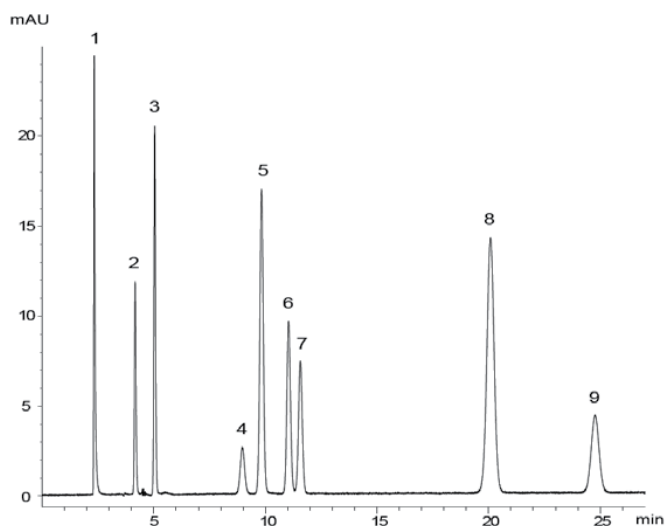
Sample preparation method



Column: YMC-Triart C18 (3 μ m, 12 nm)
50 x 2.0 mm ID
Part No.: TA12S03-0502WT
Eluent: A) acetonitrile / water / HCOOH (10/90/0.1)
B) acetonitrile / water / HCOOH (60/40/0.1)

Gradient: 5-40% B (0-12 min)
Flow rate: 0.4 mL/min
Temperature: 25 $^{\circ}$ C
Detection: UV at 254 nm
Injection: 2 μ L

Separation of water-soluble vitamins*

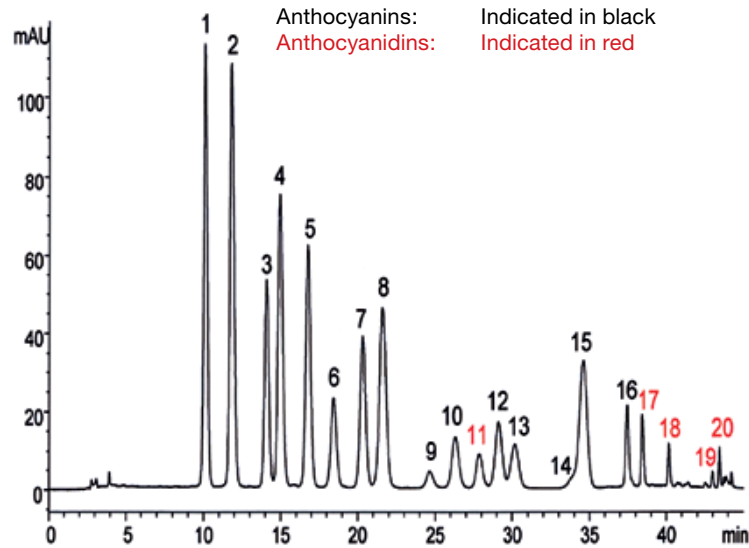


1. Thiamine HCl (Vitamin B₁)
2. Pyridoxine HCl (Vitamin B₆)
3. Nicotinamide
4. Cyanocobalamin (Vitamin B₁₂)
5. L-Ascorbic acid 2-glucoside
6. L-Ascorbic acid (Vitamin C)
7. Erythorbic acid
8. Riboflavin (Vitamin B₂)
9. Nicotinic acid

Column: YMC-Triart C18 (5 μ m, 12 nm) 250 x 4.6 mm ID
Part No.: TA12S05-2546WT
Eluent: phosphate buffer* / acetonitrile (90/10)
* Dissolve 1.4 g KH₂PO₄ in 800 mL water
→ add 26 mL 10% TBA·OH
→ adjust pH 5.2 by 20% H₃PO₄
→ add water to make 1000 mL
Flow rate: 0.8 mL/min
Temperature: 40 $^{\circ}$ C
Detection: UV at 260 nm
Injection: 10 μ L (5 μ g/mL)

Food

Analysis of anthocyanins and anthocyanidins*



Anthocyanins: Indicated in black
 Anthocyanidins: Indicated in red

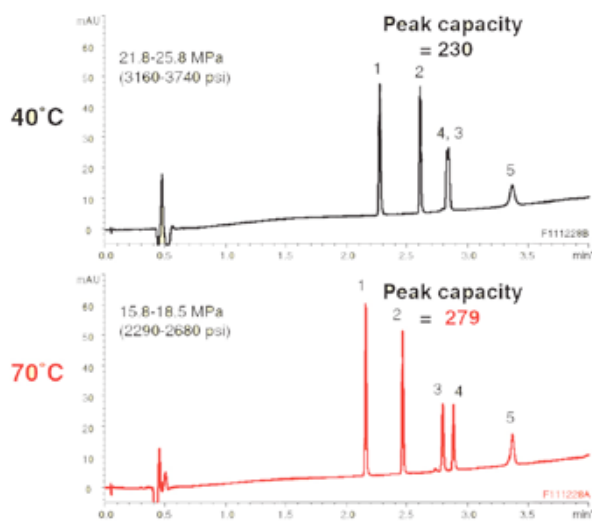
1. Delphinidin-3-O-galactoside
2. Delphinidin-3-O-glucoside
3. Cyanidin-3-O-galactoside
4. Delphinidin-3-O-arabinoside
5. Cyanidin-3-O-glucoside
6. Petunidin-3-O-galactoside
7. Cyanidin-3-O-arabinoside
8. Petunidin-3-O-glucoside
9. Peonidin-3-O-galactoside
10. Petunidin-3-O-arabinoside
11. Delphinidin
12. Peonidin-3-O-glucoside
13. Malvidin-3-O-galactoside
14. Peonidin-3-O-arabinoside
15. Malvidin-3-O-glucoside
16. Malvidin-3-O-arabinoside
17. Cyanidin
18. Petunidin
19. Peonidin
20. Malvidin

Column: YMC-Triart C18 (5 μm, 12 nm)
 250 x 4.6 mm ID
 Part No.: TA12S05-2546WT
 Eluent: A) water / formic acid (90/10)
 B) acetonitrile / methanol / water /
 formic acid (22.5/22.5/40/10)
 Gradient: 20-28% B (0-30 min),
 28-70% B (30-40 min),
 100% B (40-45 min)
 Flow rate: 1.0 mL/min
 Temperature: 25 °C
 Detection: UV/VIS at 535 nm
 Sample: commercial bilberry powder
 (1.25 mg/mL)

Peptides

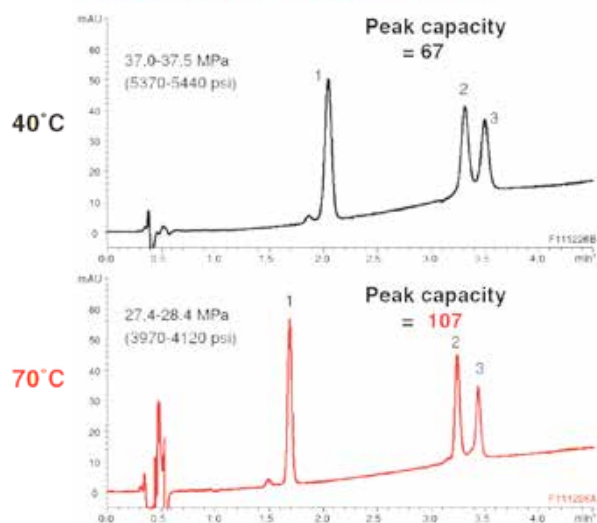
Highly efficient RP-HPLC separation of proteins and peptides using high temperature*

Mixture A (MW 500-18,400)



Analytes	MW	Peak width 1/2 (min)	
		40 °C	70 °C
Mixture A			
1. Oxytocin	1,007	0.017	0.014
2. Leu-Enkephalin	556	0.015	0.015
3. β -Endorphin	3,465	—	0.016
4. Insulin	5,733	—	0.015
5. β -Lactoglobulin A	18,400	0.043	0.030
Mixture B			
1. Lysozyme	14,300	0.069	0.044
2. α -Chymotrypsinogen	25,700	0.080	0.049
3. β -Lactoglobulin A	18,400	0.080	0.048

Mixture B (MW 14,300-25,700)



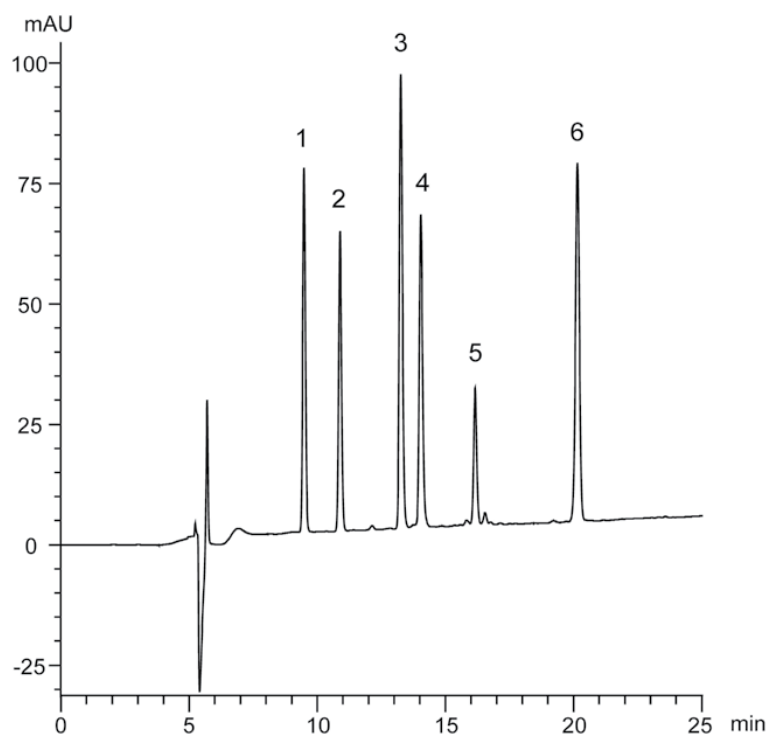
Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part-No.: TA12SP9-0502WT
 Eluent: A) water / TFA (100/0.1)
 B) acetonitrile / TFA (100/0.1) - mixture A
 B) acetonitrile / 2-propanol / TFA (50/50/0.1) - mixture B
 Gradient: 10-80% B (0-5 min) - mixture A
 30-60% B (0-5 min) - mixture B

Flow rate: 0.4 mL/min
 Detection: UV at 220 nm
 Injection: 1 μ L (50 μ g/mL) - condition A
 1 μ L (250 μ g/mL) - condition B
 System: Agilent 1200SL

PC (peak capacity) = 1 + (gradient time / peak width*)
 *peak width = $2W_{0.5h}$ average

Peptides

Peptides (MW 556 - 3,465)*



1. Oxytocin	(MW 1,007)
2. Met-Enkephalin	(MW 574)
3. Leu-Enkephalin	(MW 556)
4. Neurotensin	(MW 1,673)
5. γ -Endorphin	(MW 1,859)
6. β -Endorphin	(MW 3,465)

Column: YMC-Triart C18 (5 μ m, 12 nm)
150 x 2.0 mm ID
Part No.: TA12S05-1502WT
Eluent: A) water + 0.1% TFA
B) acetonitrile + 0.1% TFA
20-45% B (0-25 min)
Flow rate: 0.2 mL/min
Temperature: 37 $^{\circ}$ C
Detection: UV at 220 nm
Injection: 2 mL (0.075 - 0.25 mg/mL)

YMC-Triart "AQ"

YMC-Triart C18

General

The use of 100% water eluent has been a challenge in HPLC analysis for decades. Even today, many C18 materials suffer from unacceptable short lifetime, because the C18 chains collapse and this reduces the separation performance drastically. As a pioneer in this field, YMC has offered a product as early as the 80's, which presents a synonym for stability under aqueous conditions:

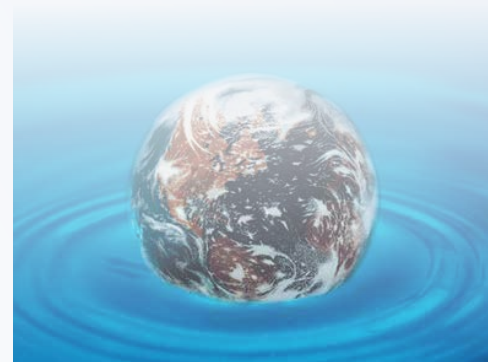
YMC-Pack ODS-AQ.

"AQ"-type phases are particularly suitable for the separation of polar substances, metabolites, pesticides, degradation products, peptides and protein digests.

1985

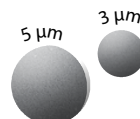


2000



YMC-Pack ODS-AQ

- "hydrophilic" C18
- balanced surface chemistry
- polar recognition
- metabolite recognition



YMC-Triart "AQ"

YMC-Triart C18

2013



Hydrosphere C18

A new, ultrapure silica base was introduced whilst adapting the surface chemistry to maintain the "AQ"-type properties.

- "hydrophilic" C18 surface for enhanced polar recognition
- stable when used with 100% aqueous eluent
- no need for ion pair reagents
- addition of 2 μm particle size for Fast-LC (YMC-Pack UltraHT)

5 μm 3 μm 2 μm

YMC-Triart "AQ" = YMC-Triart C18

The latest YMC technology platform consists of a "hybrid-style" substrate with enhanced stability against

- pH 1-12
- temp. up to 70°C
- 100% H₂O

5 μm 3 μm 1.9 μm

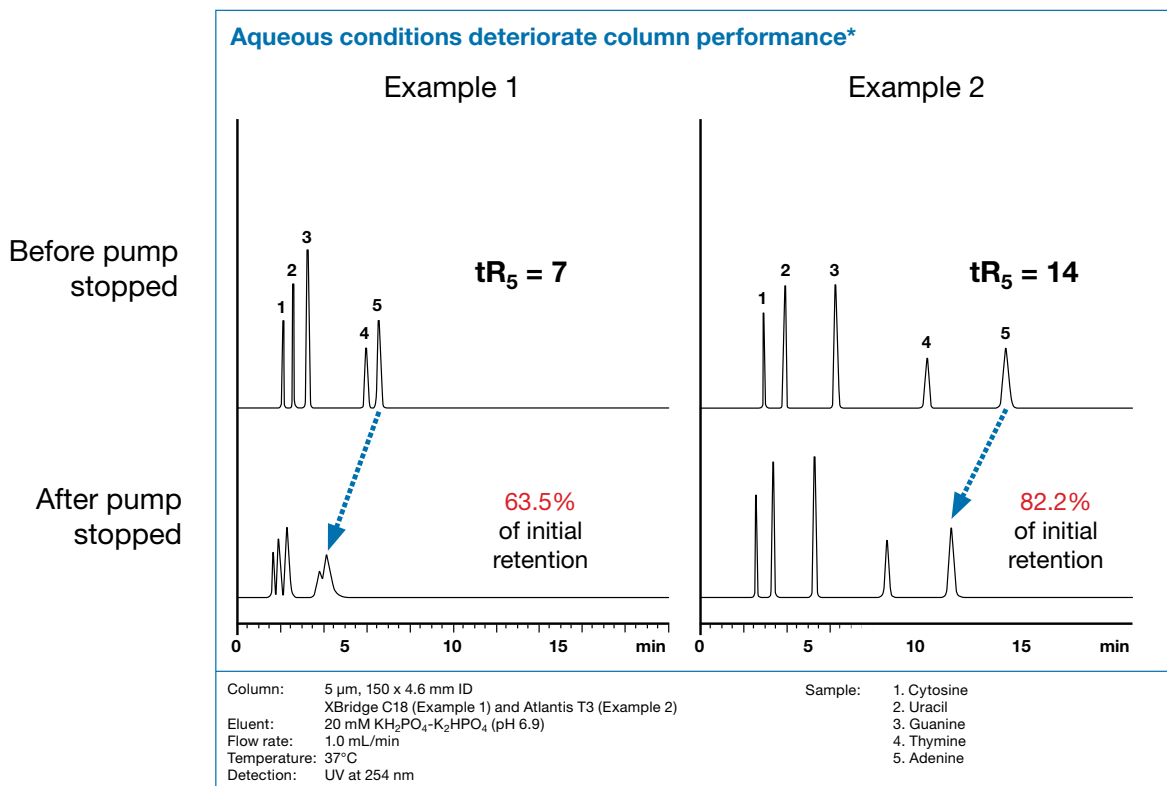
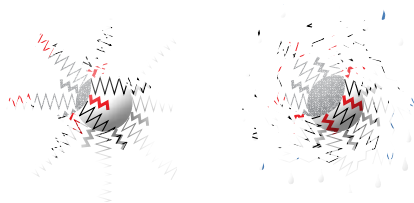


Therefore, YMC-Triart provides freedom in method development, robustness and enhanced column lifetimes for reproducible results, day after day, year after year. YMC-Triart is fully scalable within (U)HPLC \leftrightarrow HPLC with its 1.9 – 3 – 5 micron particle sizes in order to facilitate lab-to-lab method transfer.

YMC-Triart "AQ"

YMC-Triart C18

Problem with conventional C18 columns

**Why?** Image of C18 surface hydration

The columns used for applications involving 100% aqueous buffers provide shorter retention times after the flow was stopped between analyses.

This behaviour is caused by poor hydration of the phase. Polar compounds cannot easily distribute between the mobile phase and the stationary phase.

YMC-Triart "AQ"

YMC-Triart C18

Solution with YMC-Triart C18

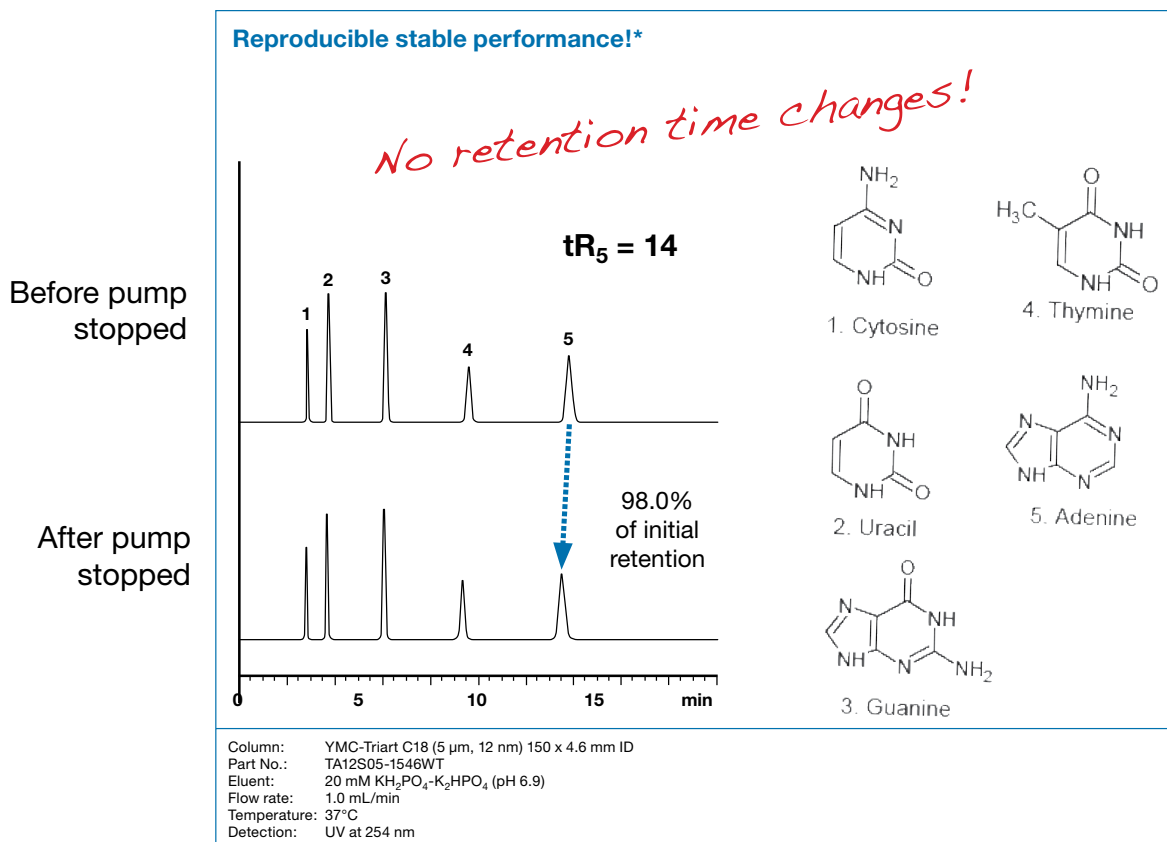
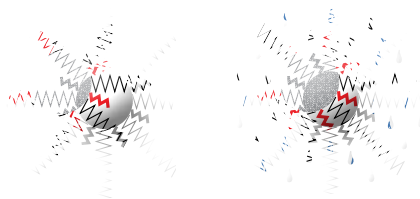


Image of C18 surface hydration

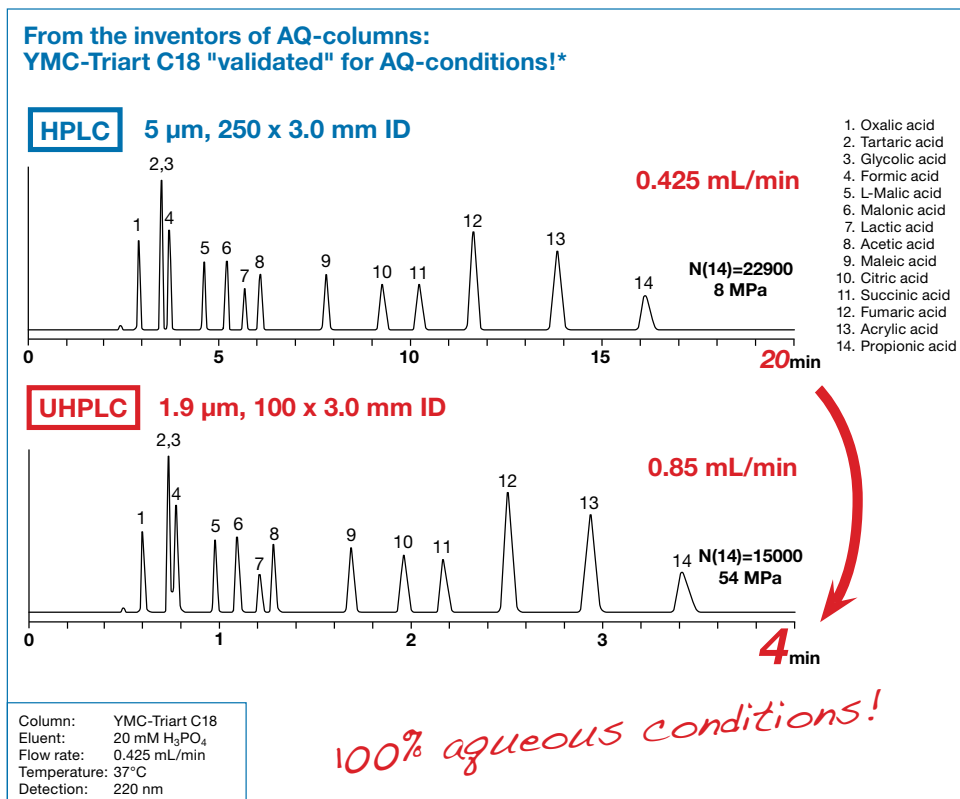


When Triart C18 columns are used for applications involving 100% aqueous buffers, the retention times are unchanged after the flow was stopped between analyses.

This is due to the improved hydration of the phase. Polar compounds can easily distribute between the mobile phase and the stationary phase.

YMC-Triart "AQ"

YMC-Triart C18

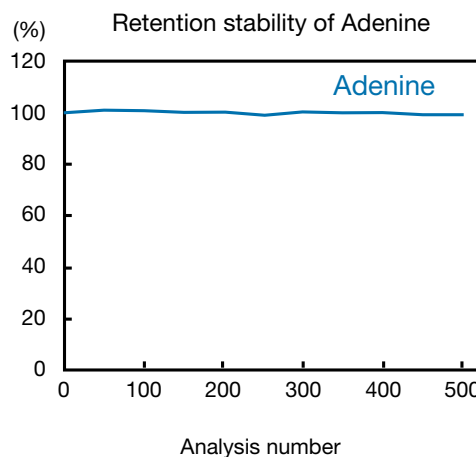
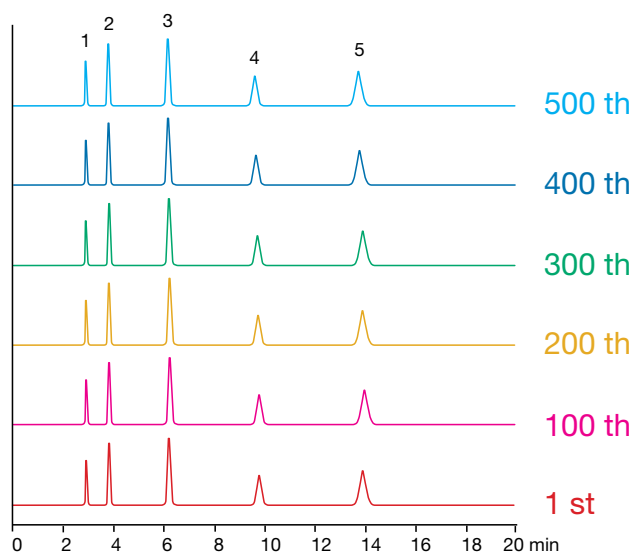


Stable under harsh conditions: pH 1-12 and temperature up to 70°C.
Stable retention times with 100% aqueous eluents!
Reproducible results day after day, column to column and lab to lab!

YMC-Triart "AQ"

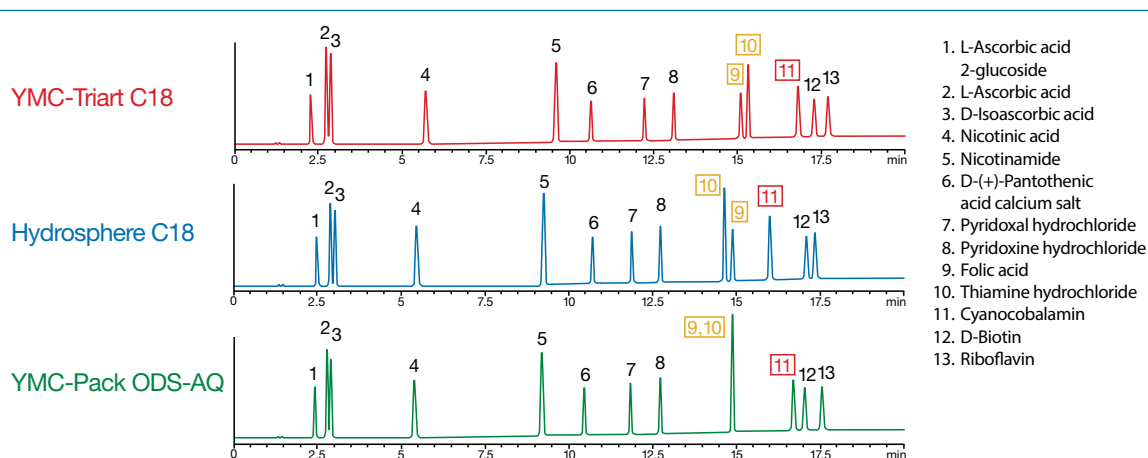
YMC-Triart C18

Proven reliability*



Column: YMC-Triart C18 (5 µm, 12 nm) 150 x 3.0 mm ID
 Part No.: TA12S05-1530WT
 Eluent: 20 mM KH₂PO₄-K₂HPO₄ (pH 6.9)
 Flow rate: 0.425 mL/min
 Temperature: 37°C
 Detection: UV at 254 nm
 Sample: 1. Cytosine 2. Uracil 3. Guanine 4. Thymine 5. Adenine

No change is found in the separation parameters including retention times, even after 500 injections when using YMC-Triart C18.



1. L-Ascorbic acid 2-glucoside
2. L-Ascorbic acid
3. D-Isoascorbic acid
4. Nicotinic acid
5. Nicotinamide
6. D-(+)-Pantothenic acid calcium salt
7. Pyridoxal hydrochloride
8. Pyridoxine hydrochloride
9. Folic acid
10. Thiamine hydrochloride
11. Cyanocobalamin
12. D-Biotin
13. Riboflavin

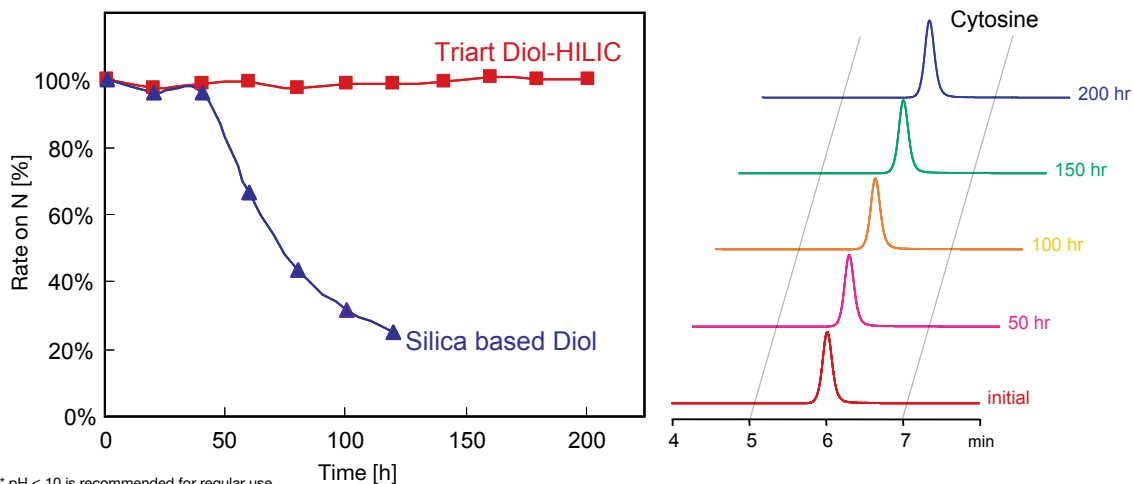
Column: 3 µm, 150 x 3.0 mm ID
 Part No.: TA12S05-1530WT
 Eluent: A) 20 mM KH₂PO₄-H₃PO₄ (pH 2.8) containing 5 mM CH₃(CH₂)₅SO₃Na
 B) 20 mM KH₂PO₄-H₃PO₄ (pH 2.8) / acetonitrile (80/20) containing 5 mM CH₃(CH₂)₅SO₃Na
 Flow rate: 0.425 mL/min
 Temperature: 40°C
 Detection: 210 nm

Retention behaviour of water-soluble vitamins on three YMC ODS phases which can be used with 100% aqueous mobile phases is compared. The retention times and peak elution order for folic acid (peak 9), thiamine hydrochloride (peak 10) and cyanocobalamin (peak 11) are different for the three phases due to the balance of hydrophobicity and hydrogen bonding capacity differing between the three phases.

HILIC

Great stability and reproducibility at high pH*

Stability in high pH (pH 11, 50 °C)**

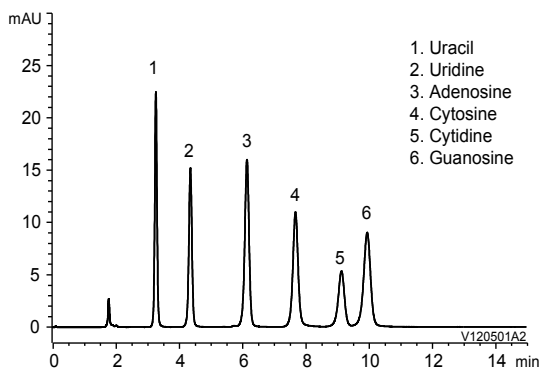


Column: 5 µm, 150 x 4.6 mm ID
Part No.: TDH12S05-1546WT
Eluent: acetonitrile / water / NH₃ (90/10/0.1) pH 11.3

Flow rate: 1.0 mL/min
Temperature: 50 °C
Sample: Cytosine

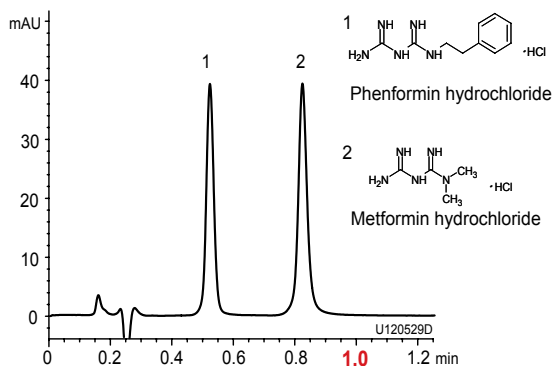
YMC-Triart Diol-HILIC offers highly reproducible separations even at high pH and high temperature. The lifetime of YMC-Triart Diol-HILIC is much longer than that of conventional silica-based Diol columns.

Nucleosides and bases*



Column: YMC-Triart Diol-HILIC (5 µm, 12 nm) 150 x 3.0 mm ID
Part No.: TDH12S05-1503WT
Eluent: 100 mM CH₃COONH₄ / acetonitrile (10/90)
Flow rate: 0.425 mL/min
Temperature: 30 °C
Detection: UV at 254 nm
Injection: 2 µL (5 ~ 10 µg/mL)

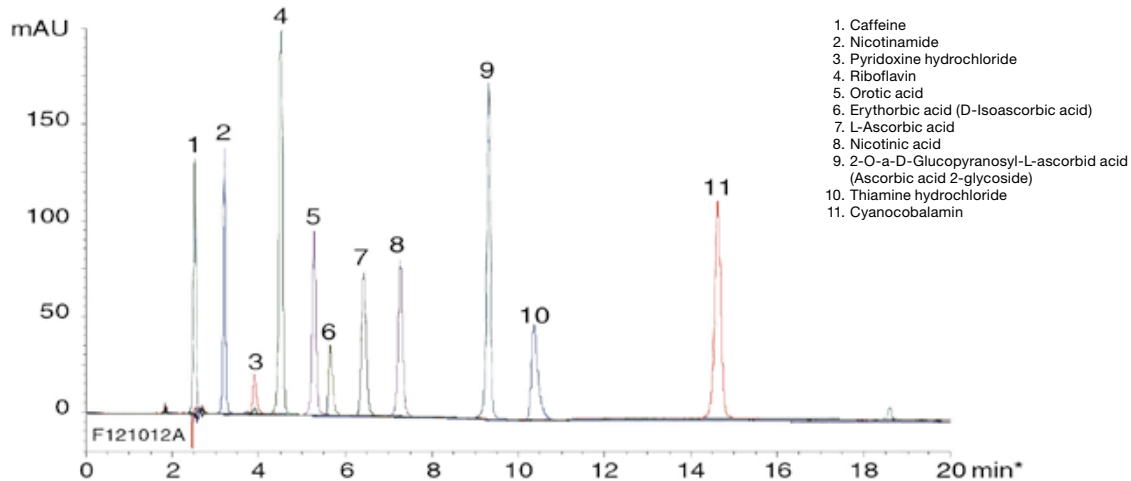
Diabetes drugs*



Column: YMC-Triart Diol-HILIC (1.9 µm, 12 nm) 50 x 2.0 mm ID
Part No.: TDH12SP9-0502PT
Eluent: 100 mM HCOOH-HCOONH₄ (pH 3.7) / acetonitrile (10/90)
Flow rate: 0.8 mL/min
Temperature: 25 °C
Detection: UV at 235 nm
Injection: 2 µL (10 µg/mL)

HILIC

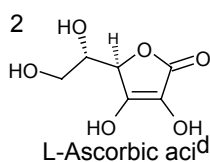
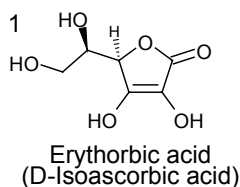
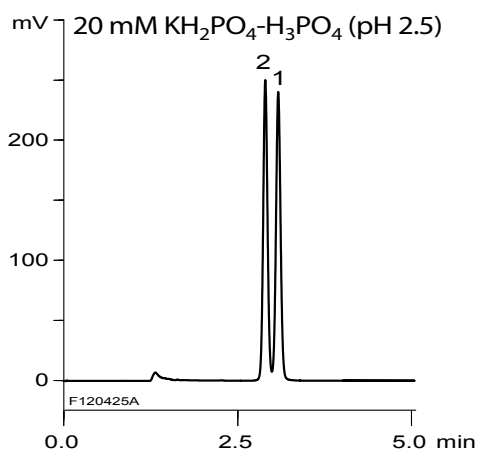
Water soluble vitamins*



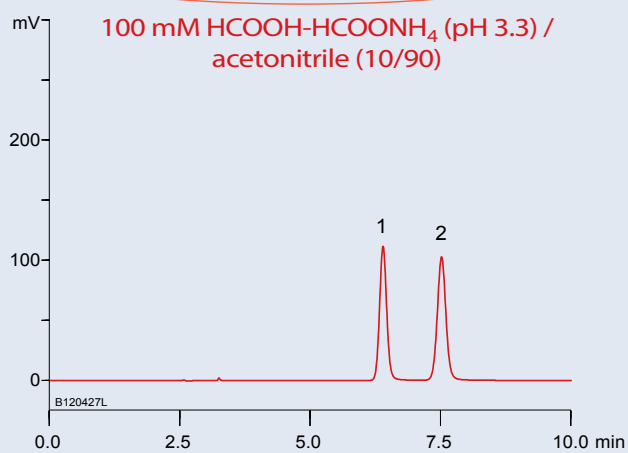
Column: YMC-Triart Diol-HILIC (5 μ m, 12 nm) 150 x 3.0 mm ID
Part No.: TDH12S05-1503PTH
Eluent: A) acetonitrile / 200 mM HCOOH-HCOONH₄ (pH 3.6) / water (90/5/5)
B) acetonitrile / 200 mM HCOOH-HCOONH₄ (pH 3.6) / water (50/5/45)
Gradient: 0-75% B (0-20 min)

Flow rate: 0.425 mL/min
Temperature: 40 °C
Detection: UV at 254 nm
Injection: 4 μ L (50 μ g/mL)

Polar and hydrophilic compounds*

RP mode
YMC-Triart C18HILIC mode
YMC-Triart Diol-HILIC

100 mM HCOOH-HCOONH₄ (pH 3.3) /
acetonitrile (10/90)



Column: 5 μ m, 150 x 3.0 mm ID
Flow rate: 0.425 mL/min
Temperature: 40 °C
Detection: UV at 254 nm
Injection: 4 μ L (0.05 mg/mL)

YMC-Triart C18 (RP) shows very weak retention and poor resolution of L-ascorbic acid and its stereoisomer (erythorbic acid) even if 100% aqueous mobile phase is used. However, YMC-Triart Diol-HILIC shows strong retention and good resolution of these compounds with mobile phase containing 90% organic solvent.

QC Data

YMC-Triart: Improved quality of particles

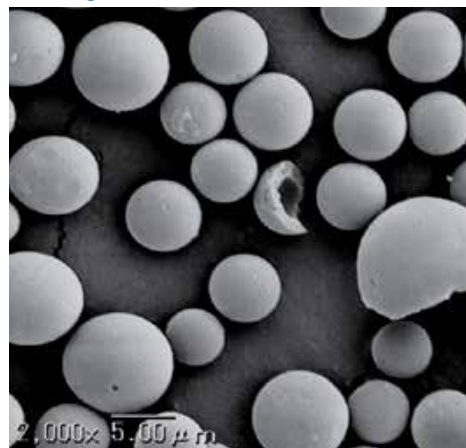
Uniform spherical particles

YMC-Triart

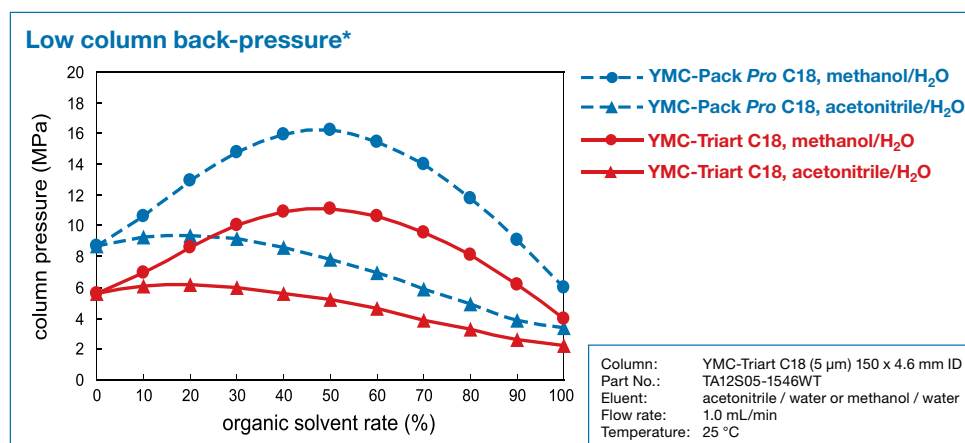


by courtesy of YMC Co., Ltd.

X-Bridge HILIC



The uniform spherical particle support is used for all YMC-Triart phases. The particles are produced using **micro-reactor** technology for the granulation process. This results in reduction of the back-pressure and leads to more reproducibility in surface modification.



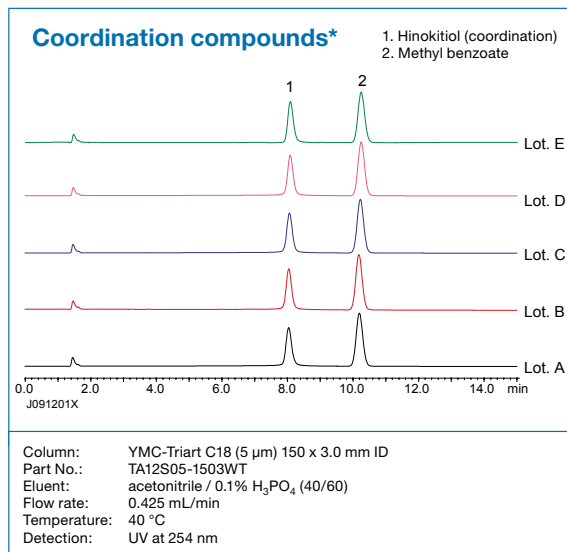
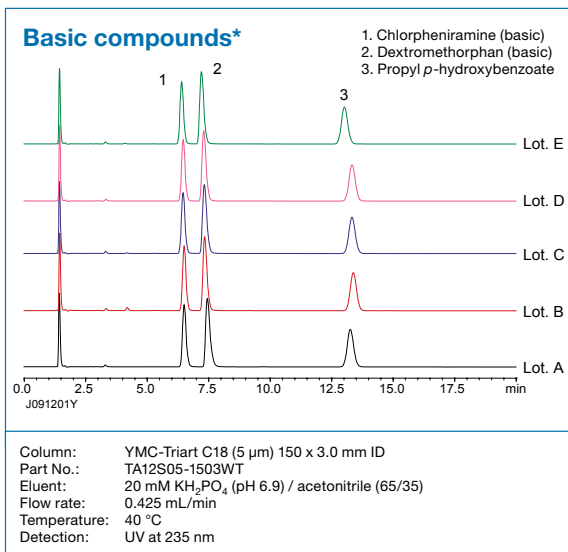
The revolutionary production technique, adapted from micro-reactor flow technology, produces a silica/organic hybrid stationary phase, with outstanding narrow pore size and particle size distributions which result in low back pressures.

YMC-Triart is designed for use under a wide range of conditions. Elution with higher viscosity methanol (compared with acetonitrile), YMC-Triart generates lower pressure (approx 30% lower than with conventional phases).

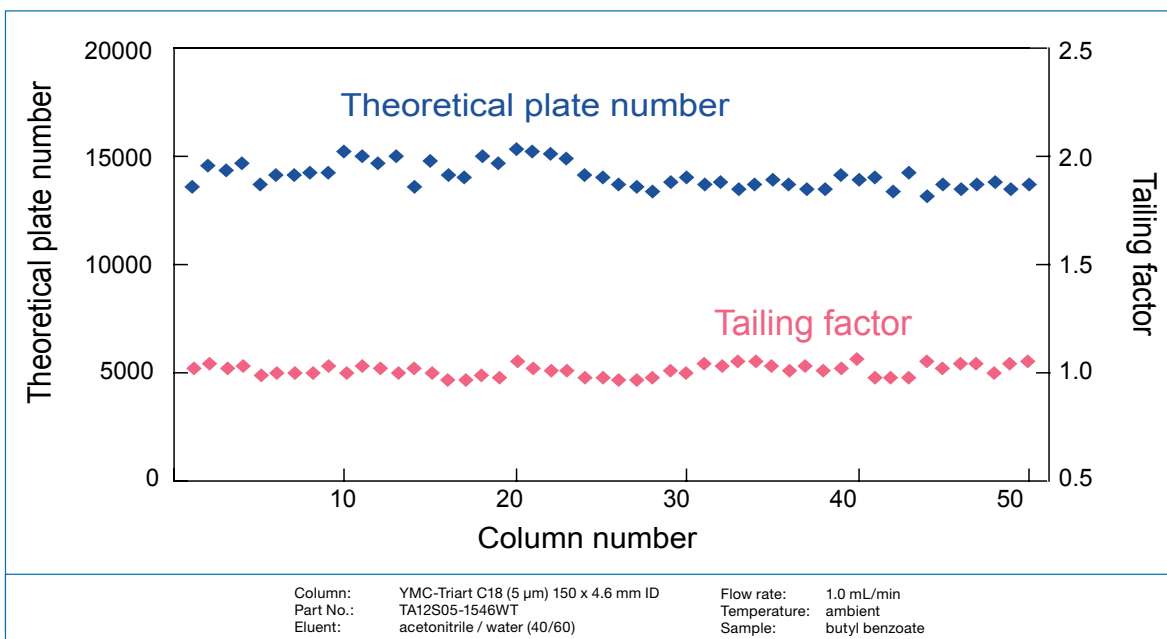
QC Data

Excellent reproducibility of YMC-Triart phases is available even for the analysis of basic and coordination compounds which normally exhibit tailing and adsorption effects.

Batch-to-batch reproducibility



The reproducibility of packed columns is shown below in terms of theoretical plate number (N) and tailing factor (Tf). YMC-Triart packed columns exhibit a very narrow range of variation.

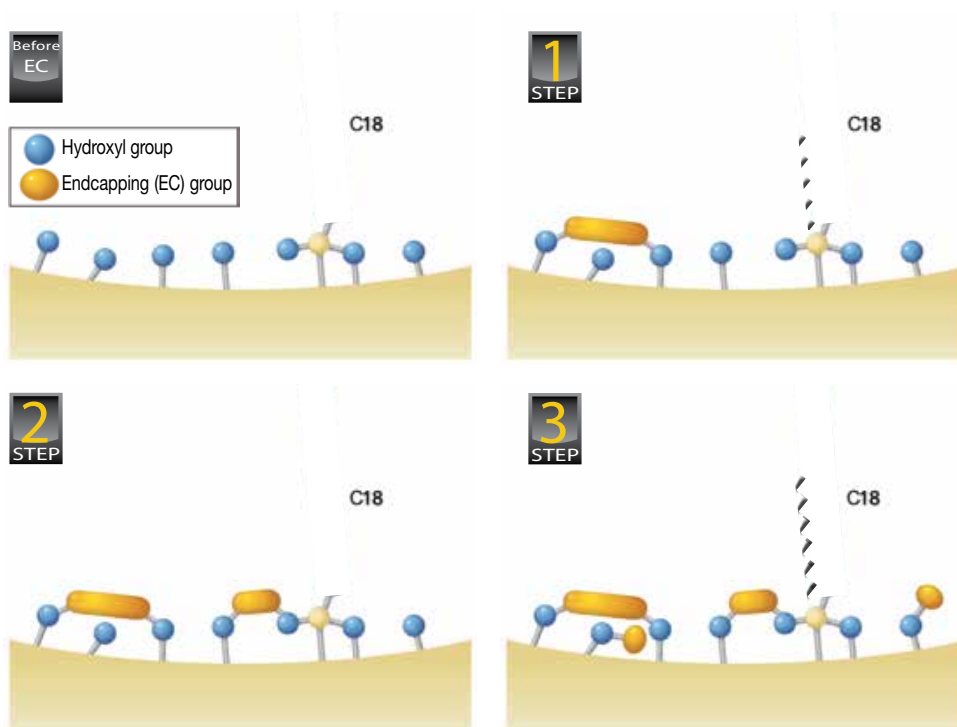


QC Data

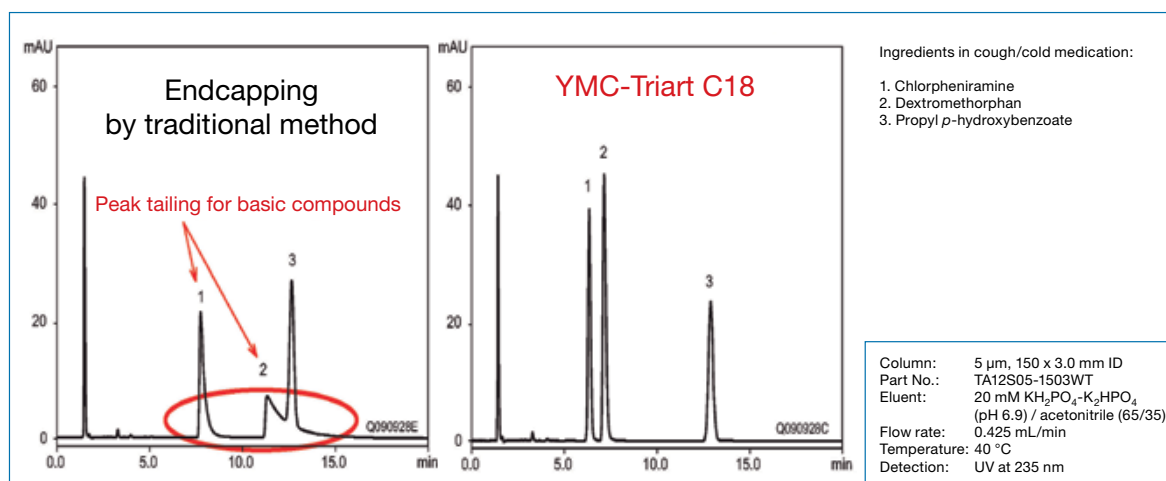
Multi-stage endcapping

After bonding the alkyl chain, there are highly reactive and less reactive silanols on the surface. In traditional bonding processes, these are reacted with a single endcapping-compound in one step. However, the highly reactive silanols can be hydrolysed easily which contributes to the poor stability. The less reactive silanols are hard to endcap which results in poor resolution due to peak tailing.

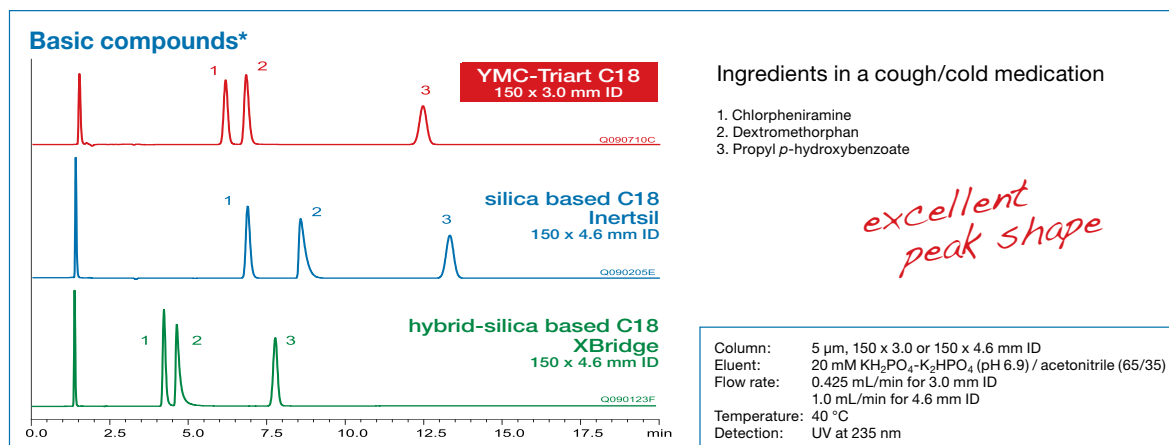
YMC-Triart phases use an innovation in endcapping called “multistage endcapping” for its surface modification process. By using a number of compounds with different reactivities in successive steps, all silanols can be capped to the maximum extent.



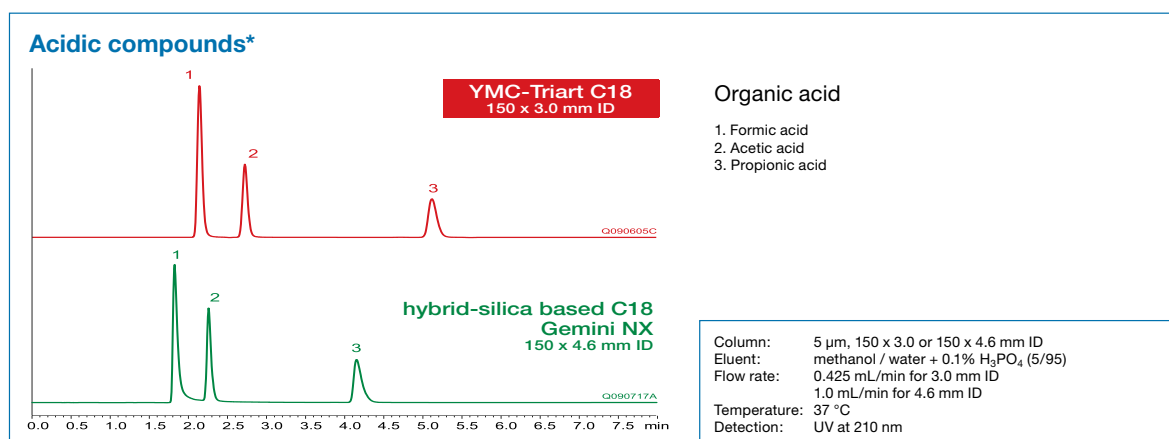
The chromatographic result of a “good” endcapping is demonstrated:



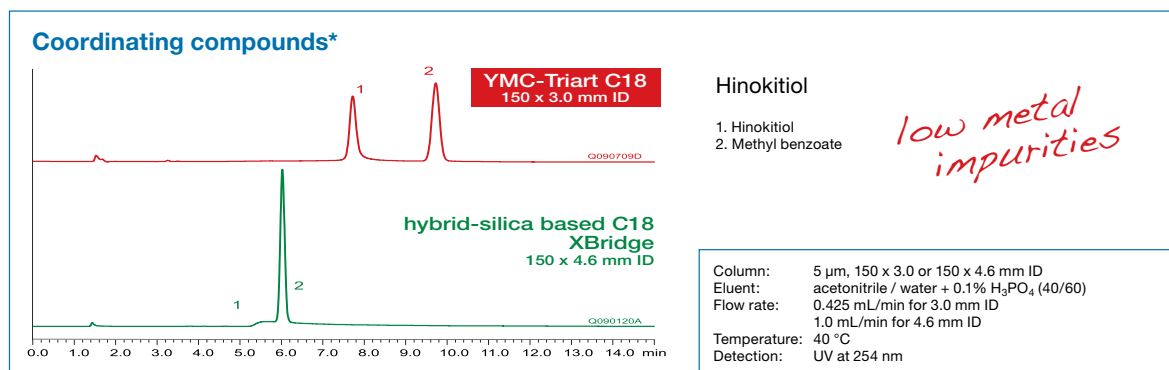
QC Data



The innovative surface modification technology results in excellent peak shapes even for basic compounds that often exhibit peak tailing with conventional silica- and hybrid silica-based reversed phase columns.



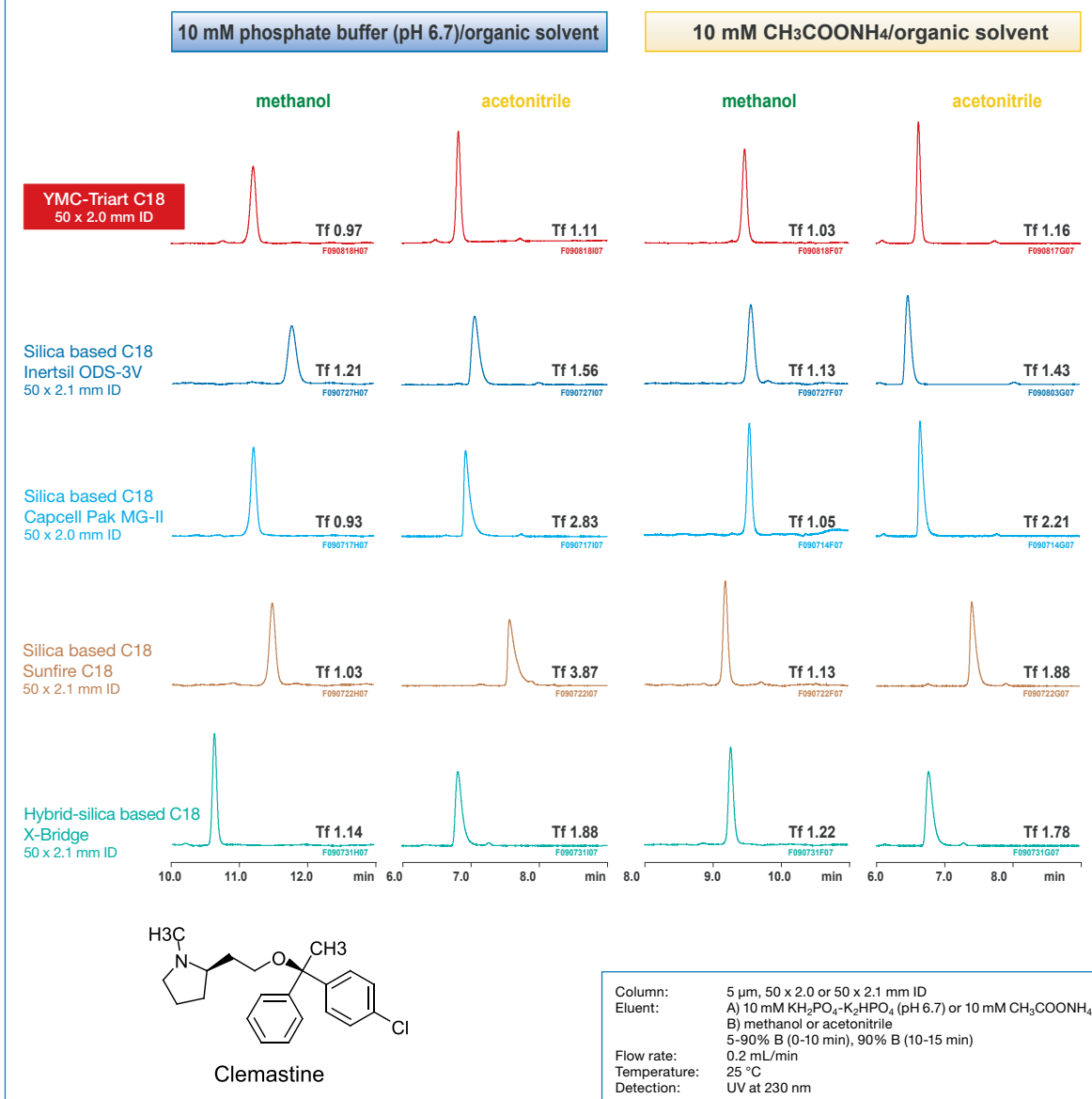
YMC-Triart phases are synthesised using methodology adapted from micro-reactor technology. This technique ensures a reduction of impurities that contribute to peak tailing during the analysis of some types acidic compounds.



YMC-Triart phases have an extremely low level of metal impurities, much lower than conventional products, ensuring excellent peak shape for coordination compounds.

QC Data

Comparison of clemastine analysis*



Clemastine is a well-known basic compound which readily exhibits peak tailing with conventional ODS columns. YMC-Triart C18 provides sharp separations with many different buffer/solvent compositions.

Ordering Information

YMC-Triart 1.9 µm UHPLC columns (max. pressure 1,000 bar)

Phase	Column ID (mm)	Column length (mm)						Guard cartridges* with 5 mm length (pack of 3)
		20	30	50	75	100	150	
C18	2.0	TA12SP9-0202PT	TA12SP9-0302PT	TA12SP9-0502PT	TA12SP9-L502PT	TA12SP9-1002PT	TA12SP9-1502PT	TA12SP9-E5Q1CC**
	2.1	TA12SP9-02Q1PT	TA12SP9-03Q1PT	TA12SP9-05Q1PT	TA12SP9-L5Q1PT	TA12SP9-10Q1PT	TA12SP9-15Q1PT	TA12SP9-E5Q1CC**
	3.0	—	—	TA12SP9-0503PT	TA12SP9-L503PT	TA12SP9-1003PT	TA12SP9-1503PT	TA12SP9-E503CC
C18 ExRS	2.0	TAR08SP9-0202PT	TAR08SP9-0302PT	TAR08SP9-0502PT	TAR08SP9-L502PT	TAR08SP9-1002PT	TAR08SP9-1502PT	TAR08SP9-E5Q1CC**
	2.1	TAR08SP9-02Q1PT	TAR08SP9-03Q1PT	TAR08SP9-05Q1PT	TAR08SP9-L5Q1PT	TAR08SP9-10Q1PT	TAR08SP9-15Q1PT	TAR08SP9-E5Q1CC**
	3.0	—	—	TAR08SP9-0503PT	TAR08SP9-L503PT	TAR08SP9-1003PT	TAR08SP9-1503PT	TAR08SP9-E503CC
C8	2.0	T012SP9-0202PT	T012SP9-0302PT	T012SP9-0502PT	T012SP9-L502PT	T012SP9-1002PT	T012SP9-1502PT	T012SP9-E5Q1CC**
	2.1	T012SP9-02Q1PT	T012SP9-03Q1PT	T012SP9-05Q1PT	T012SP9-L5Q1PT	T012SP9-10Q1PT	T012SP9-15Q1PT	T012SP9-E5Q1CC**
	3.0	—	—	T012SP9-0503PT	T012SP9-L503PT	T012SP9-1003PT	T012SP9-1503PT	T012SP9-E503CC
Phenyl	2.0	TPH12SP9-0202PT	TPH12SP9-0302PT	TPH12SP9-0502PT	TPH12SP9-L502PT	TPH12SP9-1002PT	TPH12SP9-1502PT	TPH12SP9-E5Q1CC**
	2.1	TPH12SP9-02Q1PT	TPH12SP9-03Q1PT	TPH12SP9-05Q1PT	TPH12SP9-L5Q1PT	TPH12SP9-10Q1PT	TPH12SP9-15Q1PT	TPH12SP9-E5Q1CC**
	3.0	—	—	TPH12SP9-0503PT	TPH12SP9-L503PT	TPH12SP9-1003PT	TPH12SP9-1503PT	TPH12SP9-E503CC
PFP	2.0	TPF12SP9-0202PT	TPF12SP9-0302PT	TPF12SP9-0502PT	TPF12SP9-L502PT	TPF12SP9-1002PT	TPF12SP9-1502PT	TPF12SP9-E5Q1CC**
	2.1	TPF12SP9-02Q1PT	TPF12SP9-03Q1PT	TPF12SP9-05Q1PT	TPF12SP9-L5Q1PT	TPF12SP9-10Q1PT	TPF12SP9-15Q1PT	TPF12SP9-E5Q1CC**
	3.0	—	—	TPF12SP9-0503PT	TPF12SP9-L503PT	TPF12SP9-1003PT	TPF12SP9-1503PT	TPF12SP9-E503CC
HILIC	2.0	TDH12SP9-0202PT	TDH12SP9-0302PT	TDH12SP9-0502PT	TDH12SP9-L502PT	TDH12SP9-1002PT	TDH12SP9-1502PT	TDH12SP9-E5Q1CC**
	2.1	TDH12SP9-02Q1PT	TDH12SP9-03Q1PT	TDH12SP9-05Q1PT	TDH12SP9-L5Q1PT	TDH12SP9-10Q1PT	TDH12SP9-15Q1PT	TDH12SP9-E5Q1CC**
	3.0	—	—	TDH12SP9-0503PT	TDH12SP9-L503PT	TDH12SP9-1003PT	—	—

*Guard cartridge holder required, part no. XPCUHP

**Guard cartridge: 2.1 mm ID

For other dimensions please refer to page 386-387
 For method validation and development kits refer to page 12-13

Ordering Information

YMC-Triart 3 µm high pressure rated analytical columns (max. pressure 450 bar)

Phase	Column ID (mm)	Column length (mm)							Guard cartridges* with 10 mm length (pack of 5)
		20	33	50	75	100	150	250	
C18	2.1	TA12S03-02Q1PTH	TA12S03-H3Q1PTH	TA12S03-05Q1PTH	TA12S03-L5Q1PTH	TA12S03-10Q1PTH	TA12S03-15Q1PTH	—	TA12S03-01Q1GC
	3.0	—	—	TA12S03-05Q3PTH	TA12S03-L5Q3PTH	TA12S03-10Q3PTH	TA12S03-15Q3PTH	—	TA12S03-01Q3GC
	4.6	—	TA12S03-H346PTH	TA12S03-0546PTH	TA12S03-L546PTH	TA12S03-1046PTH	TA12S03-1546PTH	TA12S03-2546PTH	TA12S03-01Q4GC
C18 ExRS	2.1	TAR08S03-02Q1PTH	TAR08S03-H3Q1PTH	TAR08S03-05Q1PTH	TAR08S03-L5Q1PTH	TAR08S03-10Q1PTH	TAR08S03-15Q1PTH	—	TAR08S03-01Q1GC
	3.0	—	—	TAR08S03-05Q3PTH	TAR08S03-L5Q3PTH	TAR08S03-10Q3PTH	TAR08S03-15Q3PTH	—	TAR08S03-01Q3GC
	4.6	—	TAR08S03-H346PTH	TAR08S03-0546PTH	TAR08S03-L546PTH	TAR08S03-1046PTH	TAR08S03-1546PTH	TAR08S03-2546PTH	TAR08S03-01Q4GC
C8	2.1	T012S03-02Q1PTH	T012S03-H3Q1PTH	T012S03-05Q1PTH	T012S03-L5Q1PTH	T012S03-10Q1PTH	T012S03-15Q1PTH	—	T012S03-01Q1GC
	3.0	—	—	T012S03-05Q3PTH	T012S03-L5Q3PTH	T012S03-10Q3PTH	T012S03-15Q3PTH	—	T012S03-01Q3GC
	4.6	—	T012S03-H346PTH	T012S03-0546PTH	T012S03-L546PTH	T012S03-1046PTH	T012S03-1546PTH	T012S03-2546PTH	T012S03-01Q4GC
Phenyl	2.1	TPH12S03-02Q1PTH	TPH12S03-H3Q1PTH	TPH12S03-05Q1PTH	TPH12S03-L5Q1PTH	TPH12S03-10Q1PTH	TPH12S03-15Q1PTH	—	TPH12S03-01Q1GC
	3.0	—	—	TPH12S03-05Q3PTH	TPH12S03-L5Q3PTH	TPH12S03-10Q3PTH	TPH12S03-15Q3PTH	—	TPH12S03-01Q3GC
	4.6	—	TPH12S03-H346PTH	TPH12S03-0546PTH	TPH12S03-L546PTH	TPH12S03-1046PTH	TPH12S03-1546PTH	TPH12S03-2546PTH	TPH12S03-01Q4GC
PPP	2.1	TPF12S03-02Q1PTH	TPF12S03-H3Q1PTH	TPF12S03-05Q1PTH	TPF12S03-L5Q1PTH	TPF12S03-10Q1PTH	TPF12S03-15Q1PTH	—	TPF12S03-01Q1GC
	3.0	—	—	TPF12S03-05Q3PTH	TPF12S03-L5Q3PTH	TPF12S03-10Q3PTH	TPF12S03-15Q3PTH	—	TPF12S03-01Q3GC
	4.6	—	TPF12S03-H346PTH	TPF12S03-0546PTH	TPF12S03-L546PTH	TPF12S03-1046PTH	TPF12S03-1546PTH	TPF12S03-2546PTH	TPF12S03-01Q4GC
HILIC	2.1	TDH12S03-02Q1PTH	TDH12S03-H3Q1PTH	TDH12S03-05Q1PTH	TDH12S03-L5Q1PTH	TDH12S03-10Q1PTH	TDH12S03-15Q1PTH	—	TDH12S03-01Q1GC
	3.0	—	—	TDH12S03-05Q3PTH	TDH12S03-L5Q3PTH	TDH12S03-10Q3PTH	TDH12S03-15Q3PTH	—	TDH12S03-01Q3GC
	4.6	—	TDH12S03-H346PTH	TDH12S03-0546PTH	TDH12S03-L546PTH	TDH12S03-1046PTH	TDH12S03-1546PTH	TDH12S03-2546PTH	TDH12S03-01Q4GC

*Guard cartridge holder required, part no. XPGCH-Q1

YMC-Triart 5 µm high pressure rated analytical columns (max. pressure 450 bar)

Phase	Column ID (mm)	Column length (mm)							Guard cartridges* with 10 mm length (pack of 5)
		20	33	50	75	100	150	250	
C18	2.1	TA12S05-02Q1PTH	TA12S05-H3Q1PTH	TA12S05-05Q1PTH	TA12S05-L5Q1PTH	TA12S05-10Q1PTH	TA12S05-15Q1PTH	—	TA12S05-01Q1GC
	3.0	—	—	TA12S05-05Q3PTH	TA12S05-L5Q3PTH	TA12S05-10Q3PTH	TA12S05-15Q3PTH	—	TA12S05-01Q3GC
	4.6	—	TA12S05-H346PTH	TA12S05-0546PTH	TA12S05-L546PTH	TA12S05-1046PTH	TA12S05-1546PTH	TA12S05-2546PTH	TA12S05-01Q4GC
C18 ExRS	2.1	TAR08S05-02Q1PTH	TAR08S05-H3Q1PTH	TAR08S05-05Q1PTH	TAR08S05-L5Q1PTH	TAR08S05-10Q1PTH	TAR08S05-15Q1PTH	—	TAR08S05-01Q1GC
	3.0	—	—	TAR08S05-05Q3PTH	TAR08S05-L5Q3PTH	TAR08S05-10Q3PTH	TAR08S05-15Q3PTH	—	TAR08S05-01Q3GC
	4.6	—	TAR08S05-H346PTH	TAR08S05-0546PTH	TAR08S05-L546PTH	TAR08S05-1046PTH	TAR08S05-1546PTH	TAR08S05-2546PTH	TAR08S05-01Q4GC
C8	2.1	T012S05-02Q1PTH	T012S05-H3Q1PTH	T012S05-05Q1PTH	T012S05-L5Q1PTH	T012S05-10Q1PTH	T012S05-15Q1PTH	—	T012S05-01Q1GC
	3.0	—	—	T012S05-05Q3PTH	T012S05-L5Q3PTH	T012S05-10Q3PTH	T012S05-15Q3PTH	—	T012S05-01Q3GC
	4.6	—	T012S05-H346PTH	T012S05-0546PTH	T012S05-L546PTH	T012S05-1046PTH	T012S05-1546PTH	T012S05-2546PTH	T012S05-01Q4GC
Phenyl	2.1	TPH12S05-02Q1PTH	TPH12S05-H3Q1PTH	TPH12S05-05Q1PTH	TPH12S05-L5Q1PTH	TPH12S05-10Q1PTH	TPH12S05-15Q1PTH	—	TPH12S05-01Q1GC
	3.0	—	—	TPH12S05-05Q3PTH	TPH12S05-L5Q3PTH	TPH12S05-10Q3PTH	TPH12S05-15Q3PTH	—	TPH12S05-01Q3GC
	4.6	—	TPH12S05-H346PTH	TPH12S05-0546PTH	TPH12S05-L546PTH	TPH12S05-1046PTH	TPH12S05-1546PTH	TPH12S05-2546PTH	TPH12S05-01Q4GC
PPP	2.1	TPF12S05-02Q1PTH	TPF12S05-H3Q1PTH	TPF12S05-05Q1PTH	TPF12S05-L5Q1PTH	TPF12S05-10Q1PTH	TPF12S05-15Q1PTH	—	TPF12S05-01Q1GC
	3.0	—	—	TPF12S05-05Q3PTH	TPF12S05-L5Q3PTH	TPF12S05-10Q3PTH	TPF12S05-15Q3PTH	—	TPF12S05-01Q3GC
	4.6	—	TPF12S05-H346PTH	TPF12S05-0546PTH	TPF12S05-L546PTH	TPF12S05-1046PTH	TPF12S05-1546PTH	TPF12S05-2546PTH	TPF12S05-01Q4GC
HILIC	2.1	TDH12S05-02Q1PTH	TDH12S05-H3Q1PTH	TDH12S05-05Q1PTH	TDH12S05-L5Q1PTH	TDH12S05-10Q1PTH	TDH12S05-15Q1PTH	—	TDH12S05-01Q1GC
	3.0	—	—	TDH12S05-05Q3PTH	TDH12S05-L5Q3PTH	TDH12S05-10Q3PTH	TDH12S05-15Q3PTH	—	TDH12S05-01Q3GC
	4.6	—	TDH12S05-H346PTH	TDH12S05-0546PTH	TDH12S05-L546PTH	TDH12S05-1046PTH	TDH12S05-1546PTH	TDH12S05-2546PTH	TDH12S05-01Q4GC

*Guard cartridge holder required, part no. XPGCH-Q1

Novel column hardware technology with increased pressure rating; specifications for the individual bonding chemistries remain identical irrespective of the column hardware format selected.

Ordering Information

YMC-Triart 3 µm analytical columns (max. pressure 200/250 bar)

Phase	Column ID (mm)	Column length (mm)							Guard cartridges* with 10 mm length (pack of 5)
		20	30	50	75	100	150	250	
C18	2.0	TA12S03-0202WT	TA12S03-0302WT	TA12S03-0502WT	TA12S03-L502WT	TA12S03-1002WT	TA12S03-1502WT	—	TA12S03-0101GC
	3.0	—	—	TA12S03-0503WT	TA12S03-L503WT	TA12S03-1003WT	TA12S03-1503WT	—	TA12S03-0103GC
	4.6	—	—	TA12S03-0546WT	TA12S03-L546WT	TA12S03-1046WT	TA12S03-1546WT	TA12S03-2546WT	TA12S03-0104GC
C8	2.0	TO12S03-0202WT	TO12S03-0302WT	TO12S03-0502WT	TO12S03-L502WT	TO12S03-1002WT	TO12S03-1502WT	—	TO12S03-0101GC
	3.0	—	—	TO12S03-0503WT	TO12S03-L503WT	TO12S03-1003WT	TO12S03-1503WT	—	TO12S03-0103GC
	4.6	—	—	TO12S03-0546WT	TO12S03-L546WT	TO12S03-1046WT	TO12S03-1546WT	TO12S03-2546WT	TO12S03-0104GC
Phenyl	2.0	TPH12S03-0202WT	TPH12S03-0302WT	TPH12S03-0502WT	TPH12S03-L502WT	TPH12S03-1002WT	TPH12S03-1502WT	—	TPH12S03-0101GC
	3.0	—	—	TPH12S03-0503WT	TPH12S03-L503WT	TPH12S03-1003WT	TPH12S03-1503WT	—	TPH12S03-0103GC
	4.6	—	—	TPH12S03-0546WT	TPH12S03-L546WT	TPH12S03-1046WT	TPH12S03-1546WT	TPH12S03-2546WT	TPH12S03-0104GC
PPP	2.0	TPF12S03-0202WT	TPF12S03-0302WT	TPF12S03-0502WT	TPF12S03-L502WT	TPF12S03-1002WT	TPF12S03-1502WT	—	TPF12S03-0101GC
	3.0	—	—	TPF12S03-0503WT	TPF12S03-L503WT	TPF12S03-1003WT	TPF12S03-1503WT	—	TPF12S03-0103GC
	4.6	—	—	TPF12S03-0546WT	TPF12S03-L546WT	TPF12S03-1046WT	TPF12S03-1546WT	TPF12S03-2546WT	TPF12S03-0104GC
HILIC	2.0	TDH12S03-0202WT	TDH12S03-0302WT	TDH12S03-0502WT	TDH12S03-L502WT	TDH12S03-1002WT	TDH12S03-1502WT	—	TDH12S03-0101GC
	3.0	—	—	TDH12S03-0503WT	TDH12S03-L503WT	TDH12S03-1003WT	TDH12S03-1503WT	—	TDH12S03-0103GC
	4.6	—	—	TDH12S03-0546WT	TDH12S03-L546WT	TDH12S03-1046WT	TDH12S03-1546WT	TDH12S03-2546WT	TDH12S03-0104GC

*Guard cartridge holder required, part no. XPGCH-Q1

YMC-Triart 5 µm analytical columns (max. pressure 200/250 bar)

Phase	Column ID (mm)	Column length (mm)							Guard cartridges* with 10 mm length (pack of 5)
		20	30	50	75	100	150	250	
C18	2.0	TA12S05-0202WT	TA12S05-0302WT	TA12S05-0502WT	TA12S05-L502WT	TA12S05-1002WT	TA12S05-1502WT	—	TA12S05-0101GC
	3.0	—	—	TA12S05-0503WT	TA12S05-L503WT	TA12S05-1003WT	TA12S05-1503WT	—	TA12S05-0103GC
	4.6	—	—	TA12S05-0546WT	TA12S05-L546WT	TA12S05-1046WT	TA12S05-1546WT	TA12S05-2546WT	TA12S05-0104GC
	10**	—	—	—	—	—	TA12S05-1510WT	TA12S05-2510WT	TA12S05-0110CC
C8	2.0	TO12S05-0202WT	TO12S05-0302WT	TO12S05-0502WT	TO12S05-L502WT	TO12S05-1002WT	TO12S05-1502WT	—	TO12S05-0101GC
	3.0	—	—	TO12S05-0503WT	TO12S05-L503WT	TO12S05-1003WT	TO12S05-1503WT	—	TO12S05-0103GC
	4.6	—	—	TO12S05-0546WT	TO12S05-L546WT	TO12S05-1046WT	TO12S05-1546WT	TO12S05-2546WT	TO12S05-0104GC
	10**	—	—	—	—	—	TO12S05-1510WT	TO12S05-2510WT	TO12S05-0110CC
Phenyl	2.0	TPH12S05-0202WT	TPH12S05-0302WT	TPH12S05-0502WT	TPH12S05-L502WT	TPH12S05-1002WT	TPH12S05-1502WT	—	TPH12S05-0101GC
	3.0	—	—	TPH12S05-0503WT	TPH12S05-L503WT	TPH12S05-1003WT	TPH12S05-1503WT	—	TPH12S05-0103GC
	4.6	—	—	TPH12S05-0546WT	TPH12S05-L546WT	TPH12S05-1046WT	TPH12S05-1546WT	TPH12S05-2546WT	TPH12S05-0104GC
	10**	—	—	—	—	—	TPH12S05-1510WT	TPH12S05-2510WT	TPH12S05-0110CC
PPP	2.0	TPF12S05-0202WT	TPF12S05-0302WT	TPF12S05-0502WT	TPF12S05-L502WT	TPF12S05-1002WT	TPF12S05-1502WT	—	TPF12S05-0101GC
	3.0	—	—	TPF12S05-0503WT	TPF12S05-L503WT	TPF12S05-1003WT	TPF12S05-1503WT	—	TPF12S05-0103GC
	4.6	—	—	TPF12S05-0546WT	TPF12S05-L546WT	TPF12S05-1046WT	TPF12S05-1546WT	TPF12S05-2546WT	TPF12S05-0104GC
	10**	—	—	—	—	—	TPF12S05-1510WT	TPF12S05-2510WT	TPF12S05-0110CC
HILIC	2.0	TDH12S05-0202WT	TDH12S05-0302WT	TDH12S05-0502WT	TDH12S05-L502WT	TDH12S05-1002WT	TDH12S05-1502WT	—	TDH12S05-0101GC
	3.0	—	—	TDH12S05-0503WT	TDH12S05-L503WT	TDH12S05-1003WT	TDH12S05-1503WT	—	TDH12S05-0103GC
	4.6	—	—	TDH12S05-0546WT	TDH12S05-L546WT	TDH12S05-1046WT	TDH12S05-1546WT	TDH12S05-2546WT	TDH12S05-0104GC

*Guard cartridge holder required, part no. XPGCH-Q1 (2.1, 3, 4 mm ID)

XPCHSPW1 (10 mm ID)

**Max. pressure 100 bar

Ordering Information

YMC-Triart, 5 µm in ACTUS high-throughput semipreparative hardware (max. pressure 300 bar)

Phase	Column ID (mm)	Column length (mm)				
		50	75	100	150	250
C18	20.0	TA12S05-0520WX	—	TA12S05-1020WX	TA12S05-1520WX	TA12S05-2520WX
	30.0	TA12S05-0530WX	TA12S05-L530WX	TA12S05-1030WX	TA12S05-1530WX	TA12S05-2530WX
C18 ExRS	20.0	TAR08S05-0520WX	—	TAR08S05-1020WX	TAR08S05-1520WX	TAR08S05-2520WX
	30.0	TAR08S05-0530WX	TAR08S05-L530WX	TAR08S05-1030WX	TAR08S05-1530WX	TAR08S05-2530WX
C8	20.0	T012S05-0520WX	—	T012S05-1020WX	T012S05-1520WX	T012S05-2520WX
	30.0	T012S05-0530WX	T012S05-L530WX	T012S05-1030WX	T012S05-1530WX	T012S05-2530WX
Phenyl	20.0	TPH12S05-0520WX	—	TPH12S05-1020WX	TPH12S05-1520WX	TPH12S05-2520WX
	30.0	TPH12S05-0530WX	TPH12S05-L530WX	TPH12S05-1030WX	TPH12S05-1530WX	TPH12S05-2530WX
PFP	20.0	TPF12S05-0520WX	—	TPF12S05-1020WX	TPF12S05-1520WX	TPF12S05-2520WX
	30.0	TPF12S05-0530WX	TPF12S05-L530WX	TPF12S05-1030WX	TPF12S05-1530WX	TPF12S05-2530WX

YMC-Triart, preparative bulk media

YMC-Triart C18-S			YMC-Triart C8-S		
Pore size (nm)	Particle size (µm)	Product Code	Pore size (nm)	Particle size (µm)	Product Code
12	10	TAS12S11	20	10	TOS20S11
	15	TAS12S16		15	TOS20S16
	20	TAS12S21		20	TOS20S21

Available in pack sizes 100 g, 500 g, 1 kg, 5 kg, 25 kg